

# Isolation and Identification of Microsporum Canis from cats in Baghdad governorate

# Marwah Abdul Hassan Bidewy<sup>1</sup>, Saleem Amin Hasso<sup>2</sup>

<sup>1,2</sup>Department of Internal & Preventive Medicine, College of Veterinary Medicine, University of Baghdad, Iraq

#### **KEYWORDS**

### Cat, Microsporum Canis, Dermatophytes, Skin Scrapings.

#### **ABSTRACT**

Microsporum canis is a keratinized skin fungus that causes diseases in animals, especially cats. It is able to infect human causes Tinea corporis and Tinea capitis . The aim of this study is to isolate and identify the dermatophyte fungi Microsporum canis from cats in Baghdad / Iraq . This research focused on the epidemiology of M. canis found in cats. A total of two hundred and eighty three hair and skin scraping samples were collected from infected cats .preliminarily examined with 10% KOH preparation, and cultured for fungal identification . For the culture; Saboruad dextrose agar was used . The results of this study showed that four samples were positive for M.canis . The pathogen was found both at the site of the lesion and at other sites in the body. The pathogen can be found in the hair of cats with and without skin lesions.

#### 1. Introduction

Dermatophytosis is the most common skin disease infecting superficial layers caused by many types of Dermatophytes (keratinophilic fungi) (Hasan, and AL-Jubori, 2015). It is zoonotic and widespread skin infection disease in pets (Boehm and Mueller, 2019). Keratinous tissues are target of dermatophytes infection as skin and hair also Claw, hooves, and horns are degraded by dermatophytes in animals (Behzadi and Ranjbar, 2014); (Mohammed, 2011) and (Minnat, 2019). Three major genera of Dermatophytes are listed; Microsporum, Trichophyton Epidermophyton based on conidial morphology of them according to Emmn's morphological classification (Rippon, 1988). The most prevalent dermatophyte agent in cats is M. canis (Aasi and Al-Aaraji., 2018) and (Saleem et al., 2020). It is very contagious, transmitted easily through direct contact and it is zoonotic in nature but not life-threatening because it can be treated (Moriello, 2014) and (Mohammed, and AL-Jibouri, 2015). The frequency of infection of *M. canis* in cats is mostly higher than those in other pet animals ,lesions of dermatophyte are etiologically correlated to this fungus (Cafarchia et al., 2006). Ringworm is commonly circular and there is hair loss, desquamation, with hair fracturing, always an erythematous edge and central curing is the usual sign lesion of M. canis in cats, Lesions is single or multiple, often on the head and face localized. Generally, lesions have been appearing on several area of the body, such as the tail and distal legs (Katiraee et al., 2016). The diagnosis was carried out through direct microscopy and cultures the specimens on each sabouraud dextrose agar (Minnat, 2019). In Iraq, there was little information about the incidence of ring worm in cats; therefore, the goal of this study is to isolate *Microsporum canis* and to assess the incidence of this species in cats.

# 2. Materials And Methods

Two hundred and eighty-three skin scraping and hair sample was collected from cats with various ages , sex and breed during the duration from April-2022 to the end of March - 2023 from different regions of Baghdad governorate .

# **Collection of samples:**

Clinically, All cats suffered from cutaneous lesions represented by loose hair, regular erythematous or just having itching. These lesions were cleaned with 70% ethyl alcohol to remove any dust and contaminated bacteria and by using the blunt edge of a sterile surgical blade, crusts and skin scales were collected by scraping from the edge of actively growing of the lesions which erythematous and inflamed margin then put onto a clean container. While in case of hair specimen collection, epilating forceps was used to pluck along the base of the hair shaft, then sealed in a sterile container; then labeled with the date of collection, age, animals name, and site of infection sex, then sent to the mycological



testing laboratory. The samples were separated into two parts: one for direct microscopic examination by using KOH 10% and other part for culturing according to (Shalaby et al., 2016).

# Diagnosis of Microsporum canis

- 1. Direct examination: The first part of each sample was treated with 10% potassium hydroxide on a clean slide then heated for 5-10 minute and let the slide to cool, and covered by cover slip for identification of fungal elements by using low powers magnification (X10) with light low intensity and lowering of the condenser. Then, use a higher magnification (X40), and higher condenser for better illumination to identify the morphology of the fungus (Kurade *et al.*, 2006) ;(Shalaby *et al.*, 2016)and (Ahmed *et al.*, 2019).
- 2. Isolation and Identification of *M. canis*: The other part of each sample was cultivated on Sabouraud dextrose agar (SDA) and incubated at 25°C for up to 2 weeks. The fungal growth was examined macroscopically and Microscopically that included large septated Macroconidia which observed by taking small part from fungal growth and mixed with one drop of lactophenol cotton blue and covered with a cover slip and examined under Microscope by using X40 lens according to (Hayyawi, 2012); (AL-Tameemi and Khalaf,2013) ;(Shalaby *et al.*, 2016) and (Jameel and Yassein,2021). An oil immersed lens (X100) was used to obtain better clarity for macroconidia of *M.canis*.

#### 3. Results and Discussion

# Macroscopic characteristic of *M. canis* infection in cats

Clinical examination of domestic cats suffering from *M. canis* was characterized clinically by circular lesions, focal and multifocal alopecia, itching, and Scaling in different area of body cats typically on the face, neck and back (**Figure 1**). On the other hand, the study showed infected hair by dermatophytosis were characterized enlarged and swollen structures with a rough and irregular surface (**Figure 2**). All fungi (*M.canis*) isolated were from male and female cats with skin lesions.

# Macroscopic Characteristic of Colonies Grown on SDA Agar

Macroscopic characteristic of *M. canis* colonies growth on SDA agar at 25°C for up to 2 weeks, white, soft and fluffy in the center with yellow or golden yellowish border closely spaced radial grooves also became white all the top with age 3-4 weeks. While reverse colony color (Undersurface view) represented by yellow that dulls to brown and darker with age as in (Figure 3)

# Microscopic Characteristics of Colonies Grown on SDA at 25C°.

Mormphology of *Microsporum canis* macroconidia on lacto phenol cotton blue stained preparations showed rough surface with knob-like end or boat like and separated into segments .Microconidia may absent or present along the length of the hyphae pyriform to round as shown in (Figure 4).

The total number of *M. canis* isolated from cats 4/283 (1.41%) as shown in (Table 1).

On the other hand, the present study demonstrated a relationship between the age of animals and infection rate with *M. canis* which represented a high percentage of infection in young age less than one year and a lower percentage in old age above one year in cats as shown in (Table 2). While the sex relation to *M. canis* infection was observed in female more than male cats with (75%) as shown in (Table 3). According to the anatomical site of lesions in cats with *M. canis*, the most sites of infection were face, neck and back as shown in (Table 4, Figure 1).



NO. of Animals	No. of M. canis isolates	Percentage
283 /cats	4	1.41%

Table 2. Percentage occurrence of *M. canis* Based on age of cats

	M. canis in Cats (n=4)	
Age	+ ve	%
< 1 year	3	75 %
≥ 1 year	1	25 %
Total	4	100 %

Table 3 . Percentage occurrence of M. can is based on sex of cats.

	M. canis in Cats (n=4)		
Sex	+ ve	%	
Female	3	75 %	
Male	1	25 %	
Total	4	100 %	

Table 4. Percentage of *M. canis* infection in cats based on Anatomical site.

Anatomical site of lesion	M. canis in Cats (n=4)	
	+ ve	%
Face	2	50 %
Neck	1	25 %
Back	1	25 %
Total	4	100 %









Figure (1): The anatomical site of lesions in cats; (A) Localized circular patch alopecia, crust, scales and redness in face female cat, 8 months old infected by *M. canis*. (B) A neck of female cat,6 months old infected with *M. canis* (C). Localized circular lesion on back of male cat,14 months old infected by *M. canis*.

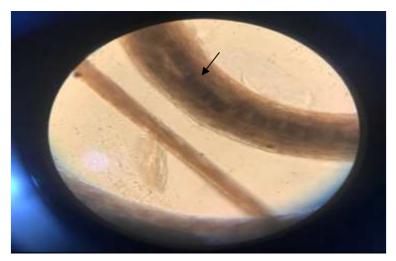


Figure (2): Infected hair by dermatophytosis (*M.canis*) were characterized enlarged and swollen structures with a rough and irregular surface (40X)

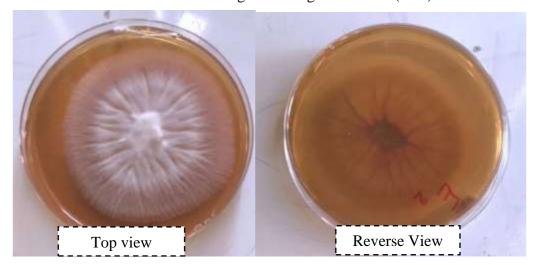


Figure (3): Macroscopic characteristic of *M. canis* colony grown on Sabouraud dextrose agar at 25°C for up to 2 weeks (Top view) and (Reverse View).



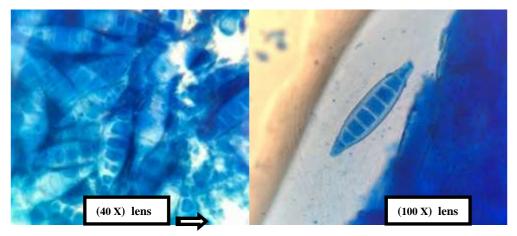


Figure (4): Macroconidia of *M. canis* stained with lactophenol cotton blue under microscope

An epidemiological study by the World Health Organization reported that there are three groups of dermatophytes that can cause infections in both humans and animals. These three groups are divided by the habitat and preferences of the fungus, are (1) anthropophilic dermatophytes, (2)zoophilic dermatophytes, and (3) geophilic dermatophytes (Chupia et al., 2022). Anthropophilic dermatophytes usually cause disease in humans; however, some species in this group can cause disease in animals, since they grow well in keratinized tissue. This allows the infection to spread from person to person via infected scabs or skin fragments. This group includes Trichophytone rubrum, Trichophytone tonsurans, Trichophytone megnini, Microsporum audouinii and Epidermophyton floccosum,. The fungi in the zoophilic dermatophyte group usually cause disease in animals but can also infect humans and cause ringworm. This group includes Microsporum canis, Microsporum equinum, Microsporum gallinae, Trichophytone verrucosum, and Trichophytone mentagrophytes. The last group is geophilic dermatophytes, which thrive in soil, plants, and environment habitats. Humans or animals can become infected by this group of fungi through contact with the environment or soils, or they may be exposed via abrasions or wounds to the skin. The members of this group include *Microsporum* gypseum, Microsporum nanum, Microsporum persicolor, and Microsporum cookei. Some studies have found that dermatophytes are dispersed across different regions of the world, with the behavior of the population in each area depending on the climate, humidity, and residential characteristics (Zhan et al., 2018). Most fungal skin diseases suffered by cats are caused by Microsporum canis (Willemse, 2015), which belongs in the group of zoophilic dermatophytes. The natural habitat of this fungus is in animals; however, this infection can also cause ringworm in humans in different parts of the body, such as the nails and head (Chupia et al., 2022). Therefore, this research aimed to detect this pathogen in cats in Baghdad, which is a pet species that lives very close to humans and may expose humans to diseases. According to this research, the percentage of infection with *M. canis* in cats in Baghdad is (1.41%). Result of M. canis isolation of this study represented by a low infection rate in cats compared to other studies. The current the study reported 1.41 % of ringworm cases were isolated from feline; this result is not consistent with a result of study conducted by (Copetti et al., 2006), which reported the isolation rate was 25.2%. The result of this study was far from (Paixão et al., 2011), who reported that M. canis was isolated from cats with (28.6%). but the investigation of (Brilhante et al., 2003), was higher slightly in which from 38 cats, 14 M.canis was isolated with (36.8%). While was higher than from the result of two studies; firstly, (Abou-Eisha et al., 2008) recorded that the dermatophytes represented by M. canis were (10%) of the examined cats. secondly (Nwiyi and Ottah,2020) revealed the isolation rate of M.canis in cats was 22%. The differentiation in the rate of isolation dermatophyte M. canis from cats between researches can be explained by the presence of virulence factors of dermatophytes spp. isolates more than others and the climate condition that is more suitable in some regions than others when these studies were conducted. These findings explain the variation allegedly occurs due to difference in relative humidity, climate, temperature, pollution of the environment and the rainfall between the geographical areas where the studies were executed; (Zenad et al., 2015). According to this research, humans who are close to and exposed to cats should be more aware of animal diseases, especially skin



diseases such as ringworm. Zookeepers, veterinarians, or people who come into contact with these animals should be aware, as contact with infected animals (both at lesions and non-lesion sites) can cause infection. Quick diagnosis leads to rapid treatment, increasing the chance of cats recovering from the disease and reducing the risk of them carrying the pathogen to humans and animals. The current diagnosis method is fungal identification from fungal culture; it takes about 10–14 days for the fungus to fully grow, so the development of a faster diagnostic method would be good for both veterinarians and zookeepers, as well as others who come into contact with animals, as it would reduce the potential risk factors (Chupia et al., 2022). Two case studies reported symptoms such as redness, dandruff, and scabs, showing severe itching (pruritic erythematous scaly plaques), and found that the symptoms of both cases were caused by M. canis. Both patients had a cat in their household. The cats were normal, strong, and healthy cats that did not show any symptoms. Preliminary examinations with Wood's lamp method provided positive results and diagnoses of M. canis, so it is possible that both of these cases were caused by *M. canis* infection from healthy cats to humans. There were no lesions, making humans think that the cats were uninfected and reducing the care taken when dealing with the animal. This increased the risk of infection. Dermatophytosis is a self-curing disease in most animals and also in cats. Infected cats should be isolated from other pets until the disease is clear (Boothe, 2012).

The findings of this study reported that animals less than one year of age were the most affected, this observation is consistent with previous studies conducted by (Minnat and Khalaf 2019). Increased susceptibility to dermatophytosis in young animals can be attributed to a several factors involving lack of previous immunity, immune system immaturity and microtrauma in skin as ectoparasites or siblings as mentioned (Moriello et al., 2017). The sex with highest susceptibility to infection with M. canis were females in cats and this result was agrees with (Nwiyi and Ottah, 2020) and the underlying reason is perhaps the greater contact of females with the contaminating animals in their living sites. The female have more interest for keeping animals such as cats in their living sites as mentioned by (Katiraee et al., 2016). Localized signs of M. canis are seen in face, neck and back but, mostly seen on face and can be attributed to the between of animals in these region with other animals carrier as mother during feeding or sleeping beside her mother (Mattei et al., 2014). The infected cat should be treated because of the zoonosis characteristic of the disease and treatment is usually recommended to shorten the course of the disease and minimize the pathogen to other susceptible animals or humans. Many antifungal drugs have been used successfully for dermatophytosis. Most commonly used antifungal drugs in veterinary medicine were ketoconazole, fluconazole, itraconazole, griseofulvin and terbinafine (Chupia et al., 2022). Besides the treatment, the prevention is very important; washing hands immediately after contact with animals (with and without skin lesion) every time is necessary when in contact with any animal. The owner should wash/change the bedding frequently because of the contaminated fomite. Mechanical removal of infected organic material/hair and surface washing, with a detergent, is the most important step for environmental cleaning/disinfection. After cleaning, a disinfectant should be used. The veterinarian should educate the owner about the necessity of treatment, control, and prevention of this disease. In Iraq, The incidence of M.canis, even if it is low, must be taken with caution because of it is possibility of some multidrug resistant strains to transmit to humans (Abulkareem Abdulshaheed, 2009).

#### 3. Conclusion

*Microsporum canis* is one of the major dermatophytic diseases in cats so; the study recommended to take high precaution toward cats due to zoonotic nature and easy transmitted of their spore.

Therefore; in this research focused on the presence of *M. canis* found in cats, collecting samples from cats with and without lesions of dermatitis. because the pathogen can be found in the hair of cats with and without skin lesions, owners, veterinary staff, and others who come into contact with the animals are at risk of infection if they are not aware or do not take precautions. The zoonotic risk and potential as an etiologic agent for a variety of diseases should be considered and investigated further.



# Reference

- [1] Aasi, S. R., and Al-Aaraji, A. M. (2018). The inhibitory effect of Trichoderma harzianum CA-07 crude extract against Trichophyton mentagrophyte and Microsporium canis. Iraqi Journal of Science, 1387-1395.
- [2] Abou-Eisha, A. M., Sobih, M. A., Hanaa, M. F., and Heba, S. E. (2008). Dermatophytes in animals and their zoonotic importance in Suez Canal area. SCVMJ, 13(2), 625-641.
- [3] Abulkareem Abdulshaheed, D. (2009). Summary Isolation and diagnosis of some fungal types from external ear canal infection cases with sensitivity test to some antifungal agents. The Iraqi Journal of Veterinary Medicine (IJVM), 33(1), 82-88
- [4] Ahmed, R. N., Mercy, B. O., and Idris, S. O. (2019). Evaluation of Secondary Metabolites of Some Fungi Isolated From Beach Soils of Lagos, Nigeria Against Some Pathogens. Iraqi Journal of Science, 2114-2122.
- [5] AL-Tameemi, H. A. A. N., and Khalaf, J. M. (2013). Isolation and identification of fungi from wounds and burns of human and farm animals. The Iraqi Journal of Veterinary Medicine, 37(2), 251-256.
- [6] Behzadi, P., Behzadi, E., and Ranjbar, R. (2014). Dermatophyte fungi: infections, diagnosis and treatment. SMU medical journal, 50-62.
- [7] Boehm, T. M., and Mueller, R. S. (2019). Dermatophytosis in dogs and cats-an update. Tierarztliche Praxis. Ausgabe K, Kleintiere/Heimtiere, 47(4), 257-268.
- [8] Boothe, D.M. (2012). Antifungal drugs. In Small Animal Clinical Pharmacology and Therapeutics, 2nd ed.; Saunders Elsevier: St. Louis, MO, USA,; pp. 368–369.
- [9] Brilhante, R. S. N., Cavalcante, C. S. P., Soares-Junior, F. A., Cordeiro, R. A., Sidrim, J. J. C., and Rocha, M. F. G. (2003). High rate of Microsporum canis feline and canine dermatophytoses in Northeast Brazil: epidemiological and diagnostic features. Mycopathologia, 156, 303-308.
- [10] Cafarchia, C., Romito, D., Capelli, G., Guillot, J., and Otranto, D. (2006). Isolation of Microsporum canis from the hair coat of pet dogs and cats belonging to owners diagnosed with M. canis tinea corporis. Veterinary dermatology, 17(5), 327-331.
- [11] Chupia, V., Ninsuwon, J., Piyarungsri, K., Sodarat, C., Prachasilchai, W., Suriyasathaporn, W., and Pikulkaew, S. (2022). Prevalence of Microsporum canis from pet cats in small animal hospitals, Chiang Mai, Thailand. Veterinary Sciences, 9(1), 21.
- [12] Copetti, M. V., Santurio, J. M., Cavalheiro, A. S., Boeck, A. A., Argenta, J. S., Aguiar, L. C., and Alves, S. H. (2006). Dermatophytes isolated from dogs and cats suspected of dermatophytosis in Southern Brazil. Acta Scientiae Veterinariae, 34(2), 119-124.
- [13] Hasan, A. M., and AL-Jubori, M. H. (2015). Determination of Optimal Temperature and pH for Radial Growth of Some Dermatophyte Species Isolated from Leukemia Patients. Iraqi journal of science, 56(1A), 95-99.
- [14] Hayyawi, S. M. (2012). Comparison of microbial isolates isolated from external ear canal of sheep and their susceptibility to antibiotics. The Iraqi Journal of Veterinary Medicine, 36(0E), 41-48.
- [15] Jameel, F. A. R., and Yassein, S. N. (2021). Virulence Potential of Penicillium Chrysogenum Isolated from Subclinical Bovine Mastitis. Iraqi Journal of Science, 2131-2142.
- [16] Katiraee, F., Asharafi Helan, J., and Teifoori, F. (2016). Multiple cases of feline dermatophytosis due to Microsporum canis transmitted to their owners. Journal of Zoonotic Diseases, 1(1), 24-30.
- [17] Kurade, S. M., Amladi, S. A., and Miskeen, A. K. (2006). Skin scraping and a potassium hydroxide mount. Indian J. Dermatol., Venereol., and Leprol., 72(3), 238.
- [18] Mattei, AS., Beber, MA., and Madrid, IM. (2014). Dermatophytosis in small animals. SOJ Microbiol. Infect. Dis. 2:1–6.
- [19] Minnat, T. R. (2019). Epidemiological, Clinical and Laboratory study of Canine Dermatophytosis in Baghdad Governorate, Iraq: Tareq Rifaaht Minnat1 and Jinan Mahmood Khalaf 2. The Iraqi Journal of Veterinary Medicine, 43(1), 183-196.
- [20] Minnat, T. R. and Khalaf, J. M. (2019). Epidemiological, Clinical and Laboratory study of Canine Dermatophytosis in Baghdad Governorate, Iraq. Iraqi J. Vet. Med. (ISSN-P: 1609-5693 ISSN-E: 2410-7409), 43(1), 183-196.

# Isolation and Identification of Microsporum Canis from cats in Baghdad governorate. SEEJPH 2024 Posted: 02-08-2024

- [21] Mohammed, S. J. (2011). A survey of dermatophytes isolated from cows and sheep in Iraq. The Iraqi journal of veterinary medicine, 35(2), 40-45.
- [22] Mohammed, S. R., and AL-Jibouri, M. H. (2015). Isolation and identification of fungi from two hospitals in Baghdad city and effect of disinfectants on some fungi. Iraqi Journal of Science, 673-682.
- [23] Moriello, K. (2014). Feline dermatophytosis: aspects pertinent to disease management in single and multiple cat situations. Journal of feline medicine and surgery, 16(5), 419-431.
- [24] Moriello, K. A., Coyner, K., Paterson, S., and Mignon, B. (2017). Diagnosis and treatment of dermatophytosis in dogs and cats. Clinical Consensus Guidelines of the World Association for Veterinary Dermatology. Veterinary Dermatology, 28(3), 266- e68.
- [25] Nwiyi, P. O., and Ottah, B. (2020). Isolation and Identification of Microsporum canis in Companion Animals from Selected Local Government Areas in Abia State. Journal of Advances in Microbiology, 9-16.
- [26] Paixão, G. C., Sidrim, J. J. C., Campos, G. M. M., Brilhante, R. S. N., and Rocha, M. F. G. (2001). Dermatophytes and saprobe fungi isolated from dogs and cats in the city of Fortaleza, Brazil. Arquivo Brasileiro de Medicina Veterinária e Zootecnia, 53, 568-573.
- [27] Rippon, J. W. (1988). Medical Mycology. WB Saunders Co, Philadelphia, USA.
- [28] Saleem, M. I., Mahmood, A. K., Qudus, A., Akbar, M., Naveed, M. T., Ahsan Nadeem, M. T., ... and ur Rehman, K. (2020). 14. Prevalence of dermatophytosis and efficacy of antifungal agents against Microsporum canis in cats. Pure and Applied Biology (PAB), 9(1), 121-131.
- [29] Shalaby, M. F. M., El-din, A. N., and El-Hamd, M. A. (2016). Isolation, identification, and in vitro antifungal susceptibility testing of dermatophytes from clinical samples at Sohag University Hospital in Egypt. Electronic physician, 8(6), 2557.
- [30] Willemse, T. (2015, April). Dermatophytosis and clinical management. In Proceedings of the 14th Chulalongkorn University Veterinary Conference (CUVC2015) (pp. 23-26).
- [31] Zenad, M. M., Badawi, N. M., and Abdul-Raheem, A. W. (2015). Cross Sectional Study on Cutaneous Mycotic Infections of Dogs and Cats in Baghdad. Vet. J, 5(4), 66-73.
- [32] Zhan, P., Dukik, K., Li, D., Sun, J., Stielow, J. B., Gerrits van den Ende, B., ... and de Hoog, G. S. (2018). Phylogeny of dermatophytes with genomic character evaluation of clinically distinct Trichophyton rubrum and T. violaceum. Studies in mycology, 89(1), 153-175.