

SEEJPH 2024 Posted: 16-08-2024

Chemical Composition Of Neem Seedlings (Azadirachta Indica L.) Sprayed With Arginine, Kinetin, And Chelated Iron

Nawras K. Mohammed¹, Kadum M. Abdullah¹, Sabah A. Fleih¹

¹Horticulture and Landscape Department, College of Agriculture, University of Kerbala, Karbala, Iraq

KEYWORDS

Active Ingredients, Amino Acids, Micronutrients, Cytokinin, Meliaceae Family

ABSTRACT

A pot experiment was carried out in the canopy of the Horticulture and Landscape Department - College of Agriculture - University of Kerbala during the 2023-2024 season to study the effect of kinetin (K) (0, 50, 100 mg L-1), arginine (A) (0, 150, 300 mg L-1), and chelated iron (F) (0 and 200 mg L-1) on some qualitative characteristics of neem seedlings. The study was implemented as a factorial experiment with a completely randomized block design with three replications. The majority of chemical features were significantly impacted by kinetin spraying, with the exception of the proportion of nitrogen, potassium, and protein in the leaves. Arginine affected all traits except total chlorophyll, while chelated iron mainly impacted potassium percentage. The research factors had varying effects on the features under investigation in the binary interaction findings. Treatment K1A1 produced the greatest rate of total chlorophyll, measuring 56.19 mg g-1 fresh weight, While the Fe2A2 treatment was superior in increasing the percentage of carbohydrates (13.31 mg g dry weight-1). With an average percentage of 3.308% for nitrogen content and 0.1588% for phosphorus, treatment K0A2 and treatment K1A2 respectively yielded the greatest percentages. However, treatment K0A1 produced a substantially higher percentage of protein, at 20.67%. Only the percentage of phosphorus was significantly affected by the triple intervention; the K1A0Fe2 treatment produced the highest rate of 0.1640%. This study concludes that High kinetin levels may have adverse effects, but when combined with nutrient-rich fertilizers, they can enhance seedling characteristics of trees with medicinal benefits.

1. Introduction

Neem trees, or Azadirachta indica L., are native to Southeast Asia, mainly Bangladesh and India. They thrive in tropical and subtropical regions and are members of the Meliaceae family (Vithalkar, 2023). This tree has garnered attention on a global scale; according to the American Academy of Sciences (Debjit Bhowmik et al., 2010), it has been dubbed the miracle tree and the tree of the twenty-first century. This tree's components are utilized in numerous industrial and agricultural applications, including the production of pharmaceuticals, since they are rich in phytochemicals including phenols, tannins, saponins, flavonoids, alkaloids, terpenoids, and fatty acids (Subapriya et al., 2005). Numerous neem active ingredients, including nimbanal, salannin, gedunin, and azadirachtin, have been demonstrated to have inhibitory functions in a variety of diseases. Many human ailments, including allergies, various skin problems, bleeding, diabetes, high blood pressure, and diuretics, have been treated with them (Chatterjee et al., 2011). The significance of neem tree leaves, branches, and roots in sustainable organic agriculture has been demonstrated by studies and research, since these parts of the tree may supply plants with natural amounts of NPK and microelements. It also contributes to the upkeep of plant health by acting as a powerful pesticide against nematodes, bacteria, fungus, and dangerous insects. For the same reason, it is also utilized as neem seed cake (Biswas et al., 2000 and Datta, 2024).

Plant hormones and growth regulators are non-food organic chemical compounds that are either intentionally supplied to or produced naturally by the plant. When introduced at different phases of growth, these chemicals alter the plant's growth and development (Paridaen, 2009; Al-Khafaji, 2014; Singh et al., 2021). Kinetin is a popular cytokinin that is widely used. It is recognized for its significant function in controlling the growth and differentiation of plant cells (Al-Doori and Hussein 2023), as well as its ability to postpone the aging process (Werner et al., 2001 and Joshi et al., 2019), promote the production of chlorophyll (Ashikari et al., 2005), and facilitate the movement and transfer of nutrients to the treated areas due to the fact that it is highly metabolic (Taiz et al., 2002), in addition to its role in the formation of some enzymes involved in the carbon fixation process (Zhang et al., 2021).



SEEJPH 2024 Posted: 16-08-2024

The building block of proteins, amino acids serve a variety of purposes in plants by controlling metabolism and the movement and storage of nitrogen. Because it starts the manufacture of proline, nitric oxide, and polyamines, arginine is the amino acid with the greatest variety of functions in living cells (Liu, 2006). It serves as the primary source of urea, and its ability to withstand environmental conditions including heat, cold, thirst, and salt is its most crucial function. Additionally, it aids in cell division, promotes the growth of roots, and produces chlorophyll (Mansour, 2000; Abdel Hafez, 2006).

Although micronutrients are not necessary for plants in large quantities, they are physiologically significant to plant life cycles. One of the elements that is essential for the respiration process is iron (Fe). It is essential for the system of many enzymes, such as catalase, peroxidas, and cytochrome, and plays a crucial role in oxidation and reduction reactions. In these reactions, iron transfers electrons, which is a crucial part of the cell's nutritional metabolism processes (Al-Mariqi, 2005). While not a component of chlorophyll, iron aids in the formation of chlorophyll. It is also vital for the metabolism of nucleic acids and chloroplasts, as well as for maintaining green matter inside plants. It contributes to the production of cytochromes, which are critical for the processes of respiration and photosynthesis (Taiz and Zeiger, 2002).

In addition to being one of the uncommon plants found across Iraq, neem trees also get little local study and research. This study intends to determine how spraying various quantities of growth regulator kinetin, amino acid arginine, and chelated iron, either separately or in combination, affects the chemical composition of neem leaves.

2. Methodology

Between the planting date of July 10, 2023, and June 1, 2024, the study was carried out under the plant canopy at the Department of Horticulture and Landscape Engineering, College of Agriculture, University of Kerbala, with 50% shade provided by saran. Neem seeds were planted in plastic bags of 5 kg of soil, which were filled with a soil mixture with peat moss at a ratio of 2 soil: 1 peat moss. The research was conducted using a factorial experiment utilizing a Randomized Complete Blocks Design (R.C.B.D.) with three replications. In each replication, there were eighteen treatments resulting from the application of three kinetin concentrations (0, 50, and 100 mg L-1), three arginine concentrations (0, 150, and 300 mg L-1), and two chelated iron concentrations (0 and 200 mg L-1). Eight plants (observations) each treatment meant that a total of 432 plants (18 treatments x 8 observations x 3 replicates) were included in the study. When the seedlings were two months old, they received four sprays in autumn and another four in the spring, applied in succession with the research factors. The percentage of nitrogen (Page et al., 1982), phosphorus (Al-Sahhaf, 1989), potassium (Hayness, 1980), and protein (Ibrahim et al., 2000) in the leaves was also determined at the conclusion of the experiment. The total chlorophyll content (mg g-1 fresh weight) was estimated using the method of Chappelle et al. (1992), and the total soluble carbohydrate content (mg g-1 dry weight) according to the method of Herbert et al. (1971).

3. Results and discussion

Total chlorophyll content in leaves (mg g-1 FW)

The results presented in Table 1 demonstrate that kinetin has a significant impact on the amount of total chlorophyll in leaves. Specifically, the concentration of 50 mg L⁻¹ produced the highest average fresh weight of 49.67 mg L⁻¹, which was not significantly different from the concentration of 100 mg L⁻¹, which produced 45.43 mg L⁻¹, while the comparison treatment produced the lowest average of 42.63 mg g⁻¹. The same table shows that iron and arginine have no discernible influence on the overall amount of chlorophyll in leaves.

147 | P a g

Chelated iron Arginine Kinetin (K) (mg L⁻¹) Iron x Arginine Chelated iron



SEEJPH 2024 Posted: 16-08-2024

		0 (K0)	50 (K1)	100 (K2)		
0 (Fe1)	0 (A0)	41.81	39.94	41.81	43.34	47.31
	150 (A1)	34.64	48.00	34.64	54.09	
	300 (A2)	51.40	46.05	51.40	44.49	
200 (Fe2)	0 (A0)	49.92	39.54	49.92	46.08	44.52
	150 (A1)	36.82	52.24	36.82	40.79	
	300 (A2)	37.81	45.94	37.81	46.68	
LSD. KxAxFe= N.S. LSD AxFe= 6.7						LSD. Fe=N.S.
Averages of Kin	etin	42.63	49.67	45.43	LSD. K=4.754	
Iron x Kinetin	0 (Fe1)	45.15	51.13	45.64		Averages of
	200 (Fe2)	40.12	48.20	45.23		Arginine
Arginine x	LSD. Fe \times K					
Kinetin	0 (A0)	40.88	48.72	44.53		44.71
	150 (A1)	40.13	56.19	45.99		47.44
	300 (A2)	46.89	44.08	45.78		45.59
	LSD. A× K	LSD. A= N.S.				

The results of the dual interaction between the study factors show that there was a significant effect on the trait mentioned above. Specifically, the intervention treatment K1A1 outperformed the comparison treatment, giving the highest rate of total chlorophyll, amounting to 56.19 mg g⁻¹, while the comparison treatment gave the lowest rate, amounting to 40.88 mg g⁻¹. Additionally, there was a significant effect from the dual interaction between arginine and chelated iron; the Fe1A1 treatment gave a rate of 54.09 mg g⁻¹, while the comparison treatment gave a rate of 43.34 mg g⁻¹. There was no discernible impact of the triple interaction of the parameters under investigation on the chlorophyll characteristic (Table 1).

Table 1 Effect of spraying with kinetin, arginine, and chelated iron on the total chlorophyll content (mg g⁻¹ fresh weight) in the leaves of neem seedlings

Total carbohydrates in leaves (mg g DW⁻¹)

The substantial impact of kinetin treatments on the leaves' total carbohydrate content is seen in Table 2. When compared to the control treatment, which recorded the lowest average of 9.23 mg g dry weight-1, the kinetin treatment with a dosage of 100 mg L-1 was superior, recording 11.33 mg g dry weight-1. This difference did not vary significantly from the treatment with a concentration of 50 mg L-1. It was evident that arginine had a major impact because at a concentration of 300 mg L-1, it produced the highest average amount of carbohydrates 11.66 mg g dry weight-1 compared to the lowest average amount 8.26 mg g dry weight-1 recorded by the comparison treatment. The aforementioned attribute was not significantly affected by iron treatments, according to the same data.

The dual interaction between the factors is also evident in Table 2, where the Fe2A2 interaction treatment outperformed the other and produced the highest rate of total carbohydrates at 13.31 mg g dry weight-1, while the Fe2A0 treatment produced the lowest rates at 7.62 mg g dry weight-1. No discernible variations exist in the total quantity of carbohydrates in the leaves of neem seedlings, as indicated by the three-way interaction among the research variables.



SEEJPH 2024 Posted: 16-08-2024

Table 2 Effect of content (mg g ⁻¹ d					the total carbohydrate	
Chelated iron	Arginine (A)	Kinetin (K) (mg L ⁻¹)		Iron x Arginine	Chelated iron	
(Fe) (mg L^{-1})	(mg L ⁻¹)	0 (K0)	50 (K1)	100 (K2)		averages
0 (Fe1)	0 (A0)	6.67	7.67	10.20	8.90	10.05
	150 (A1)	8.59	9.38	12.83	11.45	
	300 (A2)	5.99	6.49	11.51	9.80	
200 (Fe2)	0 (A0)	11.22	9.27	13.48	7.62	10.09
	150 (A1)	14.05	8.71	12.65	9.33	
	300 (A2)	8.18	10.75	13.63	13.31	
		LSD. KxA	xFe = N.S.		LSD AxFe=2.356	LSD. Fe=N.S.
Averages of Kine	tin	9.23	9.66	11.33	LSD. K=1.666	
Iron x Kinetin	0 (Fe1)	8.75	8.92	12.48		Averages of
	200 (Fe2)	9.70	10.40	10.17		Arginine
Arginine x	LSD. Fe \times K=					
Kinetin	0 (A0)	7.17	6.24	11.38		8.26
	150 (A1)	9.40	11.36	10.41		10.39
	300 (A2)	11.11	11.37	12.19		11.56
	LSD. $A \times K = 1$	N.S.				LSD. A= 1.666

Leaf content of Nitrogen %

The results presented in Table 3 indicate that the percentage of nitrogen in the leaves was not significantly affected by either kinetin or iron treatments alone. However, arginine spraying had a significant effect on this trait, as the treatment with a concentration of 150 mg L⁻¹ excelled and recorded the highest average of 2.989%, which did not differ significantly from the 300 mg L⁻¹ concentration treatment recorded 2.765%, while the comparison treatment recorded the lowest average of 2.474%.

The same data shows that the binary intervention treatments had a considerable impact since, at 3.308%, treatment K0A1 produced the greatest average nitrogen content, much higher than that of the comparative treatment, which came in at 2.438%. The preceding table indicates that the three-way interaction among the research components does not have any noteworthy impacts.

Table 3 Effect of spraying with kinetin, arginine, and chelated iron on the total Nitrogen content (%) in the leaves of neem seedlings.

Chelated iron (Fe) (mg L ⁻¹)	Arginine	Kinetin (K) (mg L ⁻¹)			Chelated iron
	(A) (mg L ⁻¹)	0 (K0)	50 (K1)	100 (K2)	Iron x Arginine	averages
	0 (A0)	2.483	2.392	3.115	2.231	
0 (Fe1)	150 (A1)	3.500	2.917	2.800	3.133	2.712
	300 (A2)	1.867	2.800	2.900	2.772	
	0 (A0)	2.117	3.225	2.915	2.717	
200 (Fe2)	150 (A1)	2.342	2.960	3.383	2.844	2.773
	300 (A2)	2.917	2.175	2.558	2.758	
		LSD. KxAx	Fe= N.S.		LSD AxFe=N.S	LSD. Fe=N.S.
Averages of Kine	tin	2.868	2.637	2.723	LSD. K=1.666	
Iron x Kinetin	0 (Fe1)	2.838	2.664	2.633		A 6
Iron x Kineun	200 (Fe2)	2.897	2.611	2.812		Averages of Arginine
	LSD. Fe \times K	= N.S				Argiiiile
	0 (A0)	2.438	2.333	2.651		2.474
Arginine x Kinetin	150 (A1)	3.308	2.508	3.150		2.989
	300 (A2)	2.858	3.070	2.367		2.765
			L	SD. $A \times K = 0$.5681	LSD. A=0.3280

SEEJPH 2024 Posted: 16-08-2024

Leaf content of Phosphorus %

The percentage of phosphorus in the leaves is shown to be significantly affected by kinetin treatments in Table 4. The treatment with a concentration of 50 mg L-1 performed best, recording the highest average of 0.1467%. This treatment did not differ significantly from the treatment with a concentration of 100 mg L-1 when compared to the control treatment, which recorded the lowest average of 0.1467% for phosphorus. 0.1338%. The table also shows that the arginine treatments had a significant impact on the characteristic in question, as evidenced by the highest average percentage of phosphorus found at a concentration of 300 mg L⁻¹, or 0.1497%, which was not significantly different from the concentration of 150 mg L⁻¹, or 0.1427%, while the comparison treatment obtained data on Phosphorus had the lowest average, at 0.1316%. The proportion of phosphorus in leaves was not significantly impacted by chelated iron applications.

Table 4. Effect of spraying with kinetin, arginine, and chelated iron on the total Phosphorous content (%) in the leaves of neem seedlings.

Chelated iron (Fe) (mg L ⁻¹)	Arginine	Kinetin (I	(mg L ⁻¹)		Iron x Arginine	Chelated iron averages	
	(A) (mg L ⁻¹)	0 (K0)	50 (K1)	100 (K2)			
	0 (A0)	0.1217	0.1190	0.1390	0.1319		
0 (Fe1)	150 (A1)	0.1413	0.1390	0.1430	0.1419	0.1409	
	300 (A2)	0.1330	0.1350	0.1483	0.1490	002.00	
	0 (A0)	0.1463	0.1640	0.1537	0.1313		
200 (Fe2)	150 (A1)	0.1410	0.1400	0.1383	0.1434	0.1417	
	300 (A2)	0.1427	0.1440	0.1547	0.1504		
		LSD. Kx	AxFe= 0.0129	7	LSD AxFe=N.S	LSD. Fe=N.S	
Averages of Kin	etin	0.1338	0.1467	0.1434	LSD. K=0.000529		
Iron x Kinetin	0 (Fe1)	0.1332	0.1484	0.1411			
non a Kineun	200 (Fe2)	0.1344	0.1450	0.1458	_	Averages of	
	LSD. Fe \times I	$\zeta = 0.00749$				Arginine	
	0 (A0)	0.1203	0.1340	0.1405		0.1316	
Arginine x Kinetin	150 (A1)	0.1402	0.1473	0.1405		0.1427	
	300 (A2)	0.1410	0.1588	0.1493		0.1497	
	LSD. A× K	LSD. $A \times K = 0.00917$					

The two-way interaction between the study factors has significant effects, as Table 4 shows. The Fe2K2 intervention treatment performed better than the comparison treatment, delivering the highest percentage of phosphorus (0.1458%) compared to 0.1332%. The K1A2 intervention treatment produced a percentage of 0.1588% compared to the comparison treatment, which resulted in the lowest percentage (0.1203%). Significant differences can be seen from the triple interaction between the factors; the Fe2K1A0 intervention treatment had the highest average percentage of phosphorus, measuring 0.1640%, while the Fe1K1A0 treatment produced the lowest averages, measuring 0.1190% (Table 4).

Leaf content of Potassium %

According to Table 5, arginine had a significant impact on the potassium content of the leaves, with the treatment at a concentration of 300 mg L-1 recording the highest average of 1.739% compared to the control treatment's lowest average of 1.481%. However, kinetin did not significantly affect the potassium content of the leaves. Chelated iron also had a substantial impact; at 200 mg L-1 of



SEEJPH 2024 Posted: 16-08-2024

concentration, it produced the greatest rate of 1.708% as opposed to the lowest rate of 1.472% for the no-spray treatment. The aforementioned attribute was not significantly affected by the two- or three-way interactions between the research components.

Table 5 Effect of spraying with kinetin, arginine, and chelated iron on the total Potassium content (%) in the leaves of neem seedlings.

Chelated iron (Fe) (mg L ⁻¹)	Arginine	Kinetin (I	K) (mg L ⁻¹)			Chelated iron	
	(A) (mg L ⁻¹)	0 (K0)	50 (K1)	100 (K2)	Iron x Arginine	averages	
	0 (A0)	2.162	1.408	1.736	1.556		
0 (Fe1)	150 (A1)	1.636	1.314	1.927	1.437	1.472	
	300 (A2)	1.164	1.661	1.094	1.423		
	0 (A0)	1.249	1.536	1.864	1.563		
200 (Fe2)	150 (A1)	1.341	1.621	1.479	1.504	1.708	
	300 (A2)	1.628	1.419	2.374	2.055	20.00	
		LSD. KxA	xFe= N.S.		LSD AxFe=N.S LSD. Fe=0.2885		
Averages of Kine	Averages of Kinetin		1.428	1.644	LSD. K=N.S.		
Iron x Kinetin	0 (Fe1)	1.737	1.265	1.413			
non a Rincin	200 (Fe2)	1.657	1.592	1.874		Averages of Arginine	
	LSD. Fe × I	K= N.S				invertiges of ringimite	
	0 (A0)	1.785	785 1.413 1.481		1.560		
Arginine x Kinetin	150 (A1)	1.686	1.172	1.554		1.471	
	300 (A2)	1.621	1.700	1.897		1.739	
	LSD. A× K	= N.S.				LSD. A=0.3533	

Leaf content of total protein %

The results presented in Table 6 indicate that the application of kinetin and chelated iron alone did not significantly impact the total protein content in the leaves. However, arginine spraying did have a significant effect on the percentage of total protein in the leaves, with the concentration of 150 mg L-1 exhibiting the highest total protein content (18.68%) in comparison to the comparison treatment, which resulted in the lowest average of 15.46% for the same characteristic.

The results of the binary interactions between the research factors demonstrated the significant impact of arginine and kinetin. Specifically, the K0A1 interaction treatment outperformed the comparison treatment, recording the highest average percentage of total protein in the leaves (20.67%), while the comparison treatment gave 15.23%. There are no discernible impacts on the percentage of total protein in the leaves, according to the three-way interaction of the research variables.

Table (6) Effect of spraying with kinetin, arginine, and chelated iron on the total Protein content (%) in the leaves of neem seedlings.

Chelated iron (Fe) (mg L ⁻¹)	Arginine	Kinetin (H	(mg L ⁻¹)			Chelated iron
	(A) (mg L ⁻¹)	0 (K0)	50 (K1)	100 (K2)	Iron x Arginine	averages
	0 (A0)	15.52	14.95	19.47	13.94	
0 (Fe1)	150 (A1)	21.87	18.23	17.50	19.58	16.95
	300 (A2)	11.67	17.50	18.12	17.33	1000
	0 (A0)	13.23	20.16	18.22	16.98	
200 (Fe2)	150 (A1)	14.64	18.50	21.15	17.78	17.33
	300 (A2)	18.23	13.59	15.99	17.24	27.00
LSD. KxAxFe= N.S.					LSD AxFe=N.S	LSD. Fe=N.S



SEEJPH 2024 Posted: 16-08-2024

Averages of Kinetin		17.02	16.48	17.92	LSD. K=N.S	
Iron x Kinetin	0 (Fe1)	16.46	16.65	17.74		
	200 (Fe2)	17.57	16.32	18.11		Averages of
	LSD. Fe \times K	K= N.S		Arginine		
	0 (A0)	15.23	14.58	16.57		15.46
Arginine x Kinetin	150 (A1)	20.67	15.68	19.69		18.68
	300 (A2)	17.86	19.19	14.79		17.28
	LSD. A× K	=3.551		LSD. A=2.050		

The distinct and combined significant effects of kinetin, arginine, and iron on various chemical properties of neem seedling leaves (total chlorophyll content, total carbohydrate content, percentage of phosphorus, nitrogen, phosphorus, potassium, and total protein) were displayed in Tables 1, 2, 3, 4, 5, and 6.

According to Al-Halabi (2012), kinetin has a significant role in extending the pace and duration of carbon absorption, which is represented by proteins, phenols, and carbohydrates. This explains why spraying kinetin improves several chemical qualities. Moreover, kinetin promotes the activity of enzymes that convert and transport carbohydrates as well as those that produce certain essential enzymes, including nitrate reductase (NR) enzymes. with relation to the process of carbon absorption (Al-Dosqi, 2008). According to Al-Shahat (2000) and Joshi et al (2019), kinetin has a crucial function in raising nitrogen build-up in old leaf sites and lowering it in new leaf sites, which promotes the synthesis of amino acids and delays the senescence of leaves. This result aligned with findings from studies conducted on olive seedlings by Al-Isaw and Al-Janabi (2021), guava (Psidium guajava) seedlings by Alkalabi and Bajlan (2023), and lettuce (Lactuca sativa L.) seedlings by Urbutis et al. (2024).

The plants that were treated with arginine exhibited superior chemical properties. This is because amino acids play a crucial role in the production of proteins and enzymes, which in turn leads to an increase in the number of leaves and overall vegetative growth. Ultimately, this efficiency boosts the process of photosynthesis, which in turn increases the amount of carbohydrates found in leaves (Hassan et al., 2014). This was also demonstrated by EL-Naggar et al. (2013) and Abd-Elkader et al. (2020).

Iron, while not being a component of chlorophyll, is crucial for its synthesis, and treating neem seedlings with chelated iron increased the amount of chlorophyll in their leaves through interactions with kinetin and arginine. The reason for this is because iron acts as an activator and catalyst, guiding a sequence of chemical events that result in the production of chlorophyll molecules, which raise the chlorophyll content of leaves (Mengel et al., 1987). These outcomes agree with the orange tree study conducted by Al-Mmureib (2008). Additionally, it was shown that neem seedlings sprayed with chelated iron combined with arginine and kinetin showed an increase in the amount of carbohydrates in the leaves. The explanation for this is that iron is involved in the formation of cytochromes, which play a crucial role in the processes of photosynthesis and respiration by accepting and transferring electrons. Additionally, iron is found in chloroplasts, which accounts for 70% of its total iron content, which makes iron essential for photosynthesis and increases the amount of carbohydrates in leaves (Al-Naimi, 2000). The outcomes of the research conducted on apple, orange, and guava trees by El-Shazly et al. (2004), Al-Mmureib (2008), and Al-Tamimi et al. (2012) are in agreement with these findings.

4. Conclusion and future scope

This study suggests that the chemical properties of tree seedlings with potential medicinal advantages can be enhanced by combining growth regulators with nutrient-rich fertilizers. The outcomes moreover shown that the combination of these variables produced more noteworthy outcomes than



SEEJPH 2024 Posted: 16-08-2024

did any one of them alone. Given certain chemical properties, it is advised against using excessive quantities of the growth regulator (Kinetin) since this might have negative effects on the plant, Considering the age of the seedling during application of the treatment.

Reference

- [1] Abdel Hafez, A. A. Al-Y. (2006). Amino acids and vitamins The use of amino acids and vitamins to improve the performance, growth and quality of horticultural crops under Egyptian conditions. United Agricultural Development (UAD).
- [2] Abd-Elkader, H. H., Massoud, H. Y., El-Baz, T. T., & El-Erian, M. A. (2020). Effect of amino acids spray on growth, flowering and keeping quality of *Gerbera jamesonii* L. as a pot plant. Journal of Plant Production, 11(2), 201-206.
- [3] Al-Desouki, H. S. A. (2008). Fundamentals of plant physiology. Rose Island Library. Mansoura Arab Republic of Egypt.
- [4] Al-Doori, M. F., & Hussein, S. A. (2023, December). Effect of Adding with Organic Nutrient (Complex-Groop) and Spraying with Kinetin on Some Characteristics of Vegetative Growth and Yield of Pomegranate Trees *Punica granatum* L. Cv. Salimi. In IOP Conference Series: Earth and Environmental Science (Vol. 1252, No. 1, p. 012079). IOP Publishing.
- [5] Al-Halabi, H. E. S. (2012). The effect of cytokinin and NPK compound fertilizer on the growth and active compounds of the black seed plant, *Nigella sativa* L. Master's thesis, Ibn Al-Haytham College of Education, University of Baghdad, Iraq.
- [6] Al-Isaw, N. A. M., & Al-Janabi, A. M. I. (2021). Effect of Foliar Application with Kinetin and Amino Acids in the Vegetative Growth and Chemical Content of Young Olive Trees cv." K18". Annals of the Romanian Society for Cell Biology, 10067-10076.
- [7] Alkalabi, C. K. J., & Bajlan, S. G. S. (2023, December). The Role of Seed Treatment with Bacterial Biofertilizer Azotobacter, Gibberellic Acid and Kinetin Before Planting in some Biochemical Characteristics in the subsequent Growth of Psidium guajava L. Seedlings. In IOP Conference Series: Earth and Environmental Science (Vol. 1262, No. 4, p. 042018). IOP Publishing.
- [8] Al-Khafaji, M. A. (2014). Plant growth regulators, their applications and horticultural uses, University House for Printing, Publishing and Translation, University of Baghdad. Ministry of Higher Education and Scientific Research, Iraq. pp. 55 348.146.p.
- [9] Al-Mmureib, K. S. A. (2008). The effect of spraying with gibberellic acid, naphthalene acetic acid, and ferrous sulphate on the growth of orange (Citrus aurantium) seedlings. Master's thesis, College of Agriculture, University of Kufa, Iraq.
- [10] Al Nuaimi, S. N. A. (2000). Principles of plant nutrition. Ministry of Higher Education and Scientific Research, Dar Al-Kutub for Printing and Publishing, University of Mosul Iraq.
- [11] Al-Sahhaf, F. H. (1989). Applied plant nutrition. Baghdad University. Ministry of Higher Education and Scientific Research. Iraq.
- [12] Al-Shahat, A. Z. N. (2000). Plant hormones and agricultural applications, Arab House for Publishing and Distribution, pp. 577, 547, 818, 283, 191.
- [13] Al-Tamimi, I. H., Al-Qatrani, N. A., and Al-Sarih, I. A. (2012). The effect of fertilization with chelated iron (FE-EDTA) on the growth of guava seedlings *Psidium guajava*, the local variety. Dhi Qar University Journal of Agricultural Research, Volume 1, Issue 2: 57-73.
- [14] Ashikari, M., Sakakibara, H., Lin, S., Yamamoto, T., Takashi, T., Nishimura, A., Angeles, E.R., Qian, Q., Kitano, H. and Matsuoka, M. (2005). Cytokinin oxidase regulates rice grain production. Science, 309(5735), 741-745.
- [15] Biswas, K., Chattopadhyay, I., Banerjee, R. K., and Bandyopadhyay, U. (2002). Biological activities and medicinal properties of neem (Azadirachta indica). Current science, 1336-1345.
- [16] Chappelle, E. W., Kim, M. S., and McMurtrey III, J. E. (1992). Ratio analysis of reflectance spectra (RARS): an algorithm for the remote estimation of the concentrations of chlorophyll a, chlorophyll b, and carotenoids in soybean



SEEJPH 2024 Posted: 16-08-2024

leaves. Remote sensing of environment, 39(3): 239-247.

- [17] Datta, H. S. (2024). Harnessing Neem (Azadirachta indica A. Juss): A Sustainable Approach to Natural Farming. International Journal of Plant & Soil Science, 36(7), 643-648.
- [18] Debjit Bhowmik, Chiranjib, Jitender Yadav, Tripathi, K. K., Sampath Kumar, K. P. (2010). Herbal Remedies of Azadirachta indica and its Medicinal Application, J. Chem. Pharm. Res. 2(1): 62-72.
- [19] El-Naggar, A. A. M., AI, A., & FEH, E. T. (2013). Response of Longiflorum X Asiatic Hybrid Lilium Plants to Foliar Spray with Some Amino Acids. Alex J. Agric. Res, 58(3), 197-208.
- [20] El-Shazly, S. M., & Dris, R. (2004). Response of 'Anna' apple trees to foliar sprays of chelated iron, manganese and zinc. J. of Food, Agriculture and Environment, 2(3), 126-130.
- [21] Gershenzon, J., & Dudareva, N. (2007). The function of terpene natural products in the natural world. Nature chemical biology, 3(7), 408-414.
- [22] Hassan, F. A., Al Taher, Z. A. A., and Saleh, A. K. N. (2014). Effect of amino acids and coconut fluid on vegetative and flowering growth and oil percentage of geranium (Pelargonium hortorum). Al-Muthanna Journal of Agricultural Sciences, Volume 3, Issue 1.
- [23] Haynes, R. J. (1980). A comparison of two modified Kjeldahl digestion techniques for multi-element plant analysis with conventional wet and dry ashing methods. Communications in Soil Science and Plant Analysis, 11(5), 459-467.
- [24] Herbert, D., Phipps, P. J., & Strange, R. E. (1971). Chapter III chemical analysis of microbial cells. In Methods in microbiology (Vol. 5, pp. 209-344). Academic press.
- [25] Ibrahim, A. M., Khalif, M. N. H., and Mustafa, I. D. (2000). Practical methods for determining chemical components in plant tissues. Alexandria Knowledge Establishment, first edition, Arab Republic of Egypt.
- [26] Joshi, S., Choukimath, A., Isenegger, D., Panozzo, J., Spangenberg, G., & Kant, S. (2019). Improved wheat growth and yield by delayed leaf senescence using developmentally regulated expression of a cytokinin biosynthesis gene. Frontiers in plant science, 10, 1285.
- [27] Liu, R., Moroi, M., Yamamoto, M., Kubota, T., Ono, T., Funatsu, A., Komatsu, H., Tsuji, T., Hara, H., Hara, H. and Nakamura, M. (2006). Presence and severity of Chlamydia pneumoniae and Cytomegalovirus infection in coronary plaques are associated with acute coronary syndromes. International Heart Journal, 47(4), 511-519.0
- [28] Mansour, M. M. F. (2000). Nitrogen containing compounds and adaptation of plants to salinity stress. Biologia plantarum, 43, 491-500.
- [29] Mengel, K., & Kirkby, E. A. (1987). Principles of plant nutrition. International Potash Institute. Berne-Worblaufen, Switzerland. (No. Ed. 4, p. 687pp).
- [30] Metwally, S. A., Abou-Leila, B. H., Taha, N. M. and Laithy, S. M. (2022). Investigate growth behavior and chemical constituents of cordyline plants under kinetin spraying. Open Access Research Journal of Life Sciences, 03(01), 033–038.
- [31] Paridan, T., and Anniekak, M. (2009). Investigating the use of plant growth regulators in New Zealand and Australia University Crops Competition New Zealand study tour project report.
- [32] Singh, V., Patel, R., Kumar, S., Pratap Sahu, M., and Ahirwal, A. (2021). Plant growth regulators and their use in plant growth and development. Agric. Environ, 2(4), 26-28.
- [33] Subapriya, R., and Nagini, S. (2005). Medicinal properties of neem leaves: a review. Current Medicinal Chemistry-Anti-Cancer Agents, 5(2), 149-156.
- [34] Taiz, L. and Zeiger, E. (2002). Plant physiology, 2nd ed. Sinauer, Sunderland.
- [35] Urbutis, M., Vaseva, I. I., Simova-Stoilova, L., Todorova, D., Pukalskas, A., & Samuolienė, G. (2024). Drought Protective Effects of Exogenous ABA and Kinetin on Lettuce: Sugar Content, Antioxidant Enzyme Activity, and Productivity. Plants, 13(12), 1641.
- [36] Vithalkar, A. (2023). The Green Gold-Neem: A Review. TIJER-INTERNATIONAL RESEARCH JOURNAL, 10(1),



SEEJPH 2024 Posted: 16-08-2024

62-77

- [37] Werner, T., Motyka, V., Strnad, M., & Schmülling, T. (2001). Regulation of plant growth by cytokinin. Proceedings of the National Academy of Sciences, 98(18), 10487-10492.
- [38] Zhang, G., Sakai, H., Yoshimoto, M., Wakatsuki, H., Tokida, T., Ikawa, H., Arai, M., Nakamura, H. and Hasegawa, T. (2021). Effect of foliar spray of kinetin on the enhancement of rice yield by elevated CO2. Journal of Agronomy and Crop Science, 207(3), 535-543.