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Single Dose of Chloroform Induces Hepatic Dysfunction with Pro-Inflammatory Cytokines Response in Vivo

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KEYWORDS

Liver Dysfunction, Pro-Inflammatory Cytokines, Chloroform

ABSTRACT

Chloroform (CH) is considered an urgent environmental pollutant that leads to critical health effects. This investigation was carried out to verify the involvement of serum cytokines in the hepatic dysfunction induced by single exposure to CH in laboratory rats. Eighteen adult male laboratory rats were separated into three groups of eight rats each. Control (CON) group included animals without any treatment, while the rest of the two groups were administrated trichloromethane at a dose of 477 mg/kg, orally then sacrificed after 1 day (CH-1) and 7day (CH-7) post-exposure. The results indicated that single-dose exposure induced liver dysfunction through significant elevation of serum activities of hepatic enzymes compared with unexposed control rats. Furthermore, chloroform intoxication caused a significant time-dependent rise in concentrations of some pro-inflammatory cytokines in serum of exposed rats. In conclusion, a single high dose of chloroform causes liver dysfunction and promotes the response of pro-inflammatory cytokines in rats.

1. Introduction

Chloroform (CH), also called trichloromethane (CHCl3), is a highly volatile organic compound that is an uncolored liquid with a distinctive odour [1]. It was introduced in 1847 as a powerful anesthetic, until its clinical usage declined in 1976 due to its adverse side effects [2]. Inhalation of CH induces central nervous system depression, leading to anesthesia, and can be fatal at high concentrations [3]. It is a common pollutant in atmosphere and water, and released into air as a by-product of its formation in chlorination of drink water, sewage, and swimming pools [4,5]. Its volatility allows it to go from water to the atmosphere, where it remains for several months before being removed through chemical conversion [6,7]. It is also a by-product of paper bleaching and emissions from vehicle exhaust [8]. It is worth noting that CH is classified as a hazardous substance on the list of the most important priority substances by agency for toxic substances and disease registry. In addition, it has been considered a potential carcinogen [9]. In sequence, liver, kidneys, and central nervous system are the target organs for CH poisoning [10]. Both humans and animals get identical poisoning symptoms from it; the average oral fatal dose for humans is roughly 45 g. [11]. Mammals are generally good absorbers, metabolizers, and excretors of chloroform following exposure, and the bloodstream carries the chemical throughout the body extensively [12].

The main byproduct of the metabolism of chloroform is carbon dioxide (CO2), which is mostly expelled by the lungs. However, some of the CO2 is integrated into endogenous metabolites and may be expelled as various amino acids, urea, bicarbonate, and methionine [13]. It is extremely harmful to the liver, albeit the first 12 to 48 hours following exposure may not show full signs of damage [14]. Acute heart failure, lung failure, and centrilobular liver necrosis can all be fatal outcomes of acute exposure to excessive doses of CH. [15]. It is well established that liver is the key site of metabolism of xenobiotics and hepatotoxicity resulting from exposure to organic solvents is rarely diagnosed or even suspected [16,17]. There is strong suspicion that chloroform metabolites may have mutagenic and immune toxic effects [18,19]. It is requisite to investigate the role of inflammatory mediators including cytokine response, in the previously reported hepatic injury induced by high dose chloroform in vivo [20,21]. Thus, this experimental study aimed to assess contribution of serum cytokines to liver injury in a rat model caused by a single chloroform treatment.

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Animals and treatments

For this experiment, a total of eighteen fully grown male Wistar rats with weigh ranged around 180 - 220 g. were selected from the animal resources center. They were housed in appropriate clean cages and allowed to acclimatize to standard laboratory environmental conditions in terms of temperature, lighting and humidity for one week prior starting the experiment. They were maintained on a standard diet with easy access to water. All animals were treated ethically according to the National Institutes of Health guidelines (1978). Randomly, they were set into three groups, each consisting of six rats. Following receiving a single oral dosage of 477 mg/kg of trichloromethane [22], the first group (CON) was kept as a control without receiving any treatment. The other two groups were euthanized one day (CH-1) and seven days (CH-7) following the exposure. Blood samples from cardiac punctures were taken, put in designated tubes, processed to extract the serum, and then kept cold at–40 ° C until analysis.

Assessment of liver enzymes and proinflammatory cytokines

A standardized ELISA methodology was used to measure liver enzymes levels including ALT & AST, ALP, and GGT in serum samples. As directed by the kit, standards were made by serial dilution. Serum concentrations of the pro-inflammatory cytokines including IL-1 β , IL-6 and TNF- α were evaluated using commercially standardized ELISA protocol. According to industry recommendations, serum samples were processed.

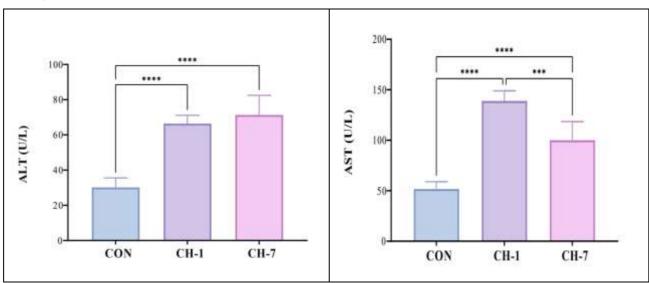
Data analysis

The SPSS (version 25) software was applied to process all results, and $M\pm$ SD was the output. Differences between research groups were determined using one-way analysis of variance (ANOVA), followed by Tukey's post hoc analysis interpretation. A significance level set at p < 0.05 was used. GraphPad Prism was utilized to design the graphs.

Results

Effect on liver enzymes levels

The detrimental effect of chloroform on the serological levels of studied rats after 1 day and 7 days of CH exposure is represented in Fig. (1). The findings confirmed that CH-treatment led to a remarkable increasing in levels of analyzed liver enzymes in serum of CH-1 and CH-7 rats compared to controls. It is worth noting that this increase was gradual and time-dependent in enzymes (ALT, ALP, and GGT) in CH-treated rats.





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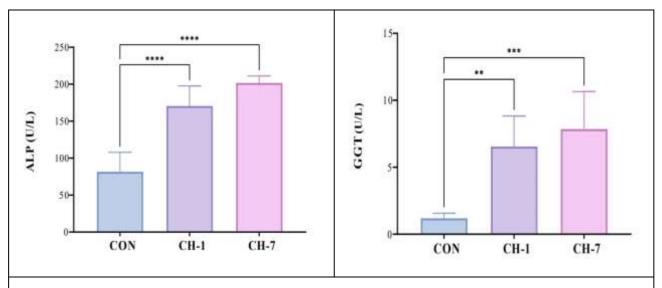
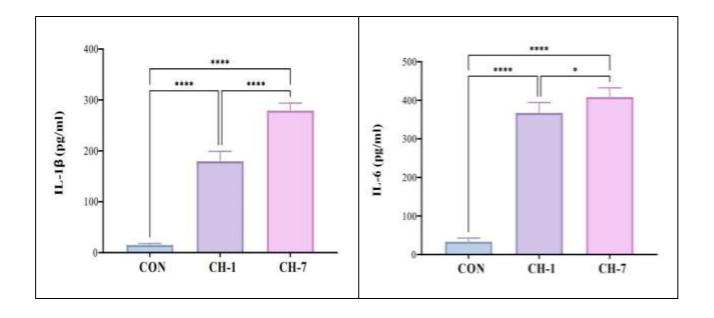


Figure 2: Impact of chloroform on serum concentrations of liver enzymes in three experiment groups. **: significant at (p < 0.01), *** (p < 0.001); **** (p < 0.0001).

Effect on proinflammatory cytokines levels

Results confirmed that exposure to chloroform induced a significant time-dependent increase (p < 0.0001) in the serum levels of IL-1 β , IL-6, and TNF- α in CH-1 and CH-7 rats compared to control (CON) group as shown in Fig. (2).





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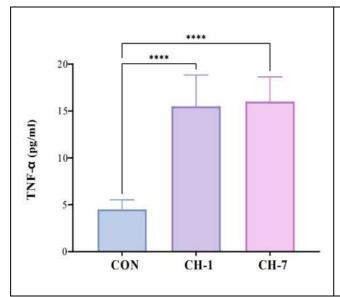


Figure 2: Impact of chloroform on serum concentrations of pro-inflammatory cytokines in three experiment groups. *: significant at (p < 0.05); **** (p< 0.0001).

Discussion

The liver is a vital organ involved in extramedullary hematopoiesis, has a high regenerative capacity, and is essential for detoxification and metabolism [23,24]. In the present era, the liver damage caused by environmental chemical pollutants has become overwhelming due to continued exposure, thus increasing the risk of poisoning [25-27]. Serum levels of these liver enzymes are measured during liver function testing, which is proven to be a useful method for assessing the liver's health [28]. Thus, a rise in these serum enzyme levels could be a sign of liver damage or inflammation [29]. According to the results, a single high dose of chloroform induced liver damage through disruption levels of hepatic functional indices in exposed rats. The levels of liver enzymes were significantly raised compared to control group, this indicated that chloroform increased cell membrane permeability and induced hepatocytes damage [30]. Our findings were consistent with a previous study by Chima et al. as they found that chloroform-intoxicated albino rats (100 mg/kg/b.wt) had induced activity of serum hepatic markers ALT, ALP, AST and GGT [31]. Furthermore, Okechukwu et al confirmed that experimental rats intoxicated with chloroform demonstrated a substantial increase (p < 0.05) in serum levels of ALT, ALP, and AST, suggesting that the treated animals were under oxidative stress. They explained this by the disruption of the plasma membrane and damage to hepatocellular cells, which resulted in rapid release of these enzymes into the bloodstream [32]. In a different study, Somade et al. found that oral trichloromethane exposure to rats at a level of 200 mg/kg induced hepatotoxicity, as evidenced by a discernible rise in the liver's expression of nitric oxide, H2O2, apoptotic cells, nuclear factor kappa B, and oxidative stress indicator. Additionally, p53 expression and antioxidant activities were clearly reduction than in the unexposed group [33]. The hepatotoxicity of chloroform has been reported to be due to phosgene-mediated depletion of cellular glutathione or increased amounts of covalent binding to hepatic macromolecules, such as proteins, DNA, and lipids, resulting in cellular malfunction and death [34]. Primarily, CH is metabolized in the liver through oxidation-reduction pathways, with the major oxidation products being carbon dioxide and reactive metabolites containing phosgene and trichloromethyl radicals. [35,36]. Oxygen tension, species, tissue, and dose all affect the balance of the pathways, which are based on cytochrome P450-dependent enzyme activation [37]. The results also proved the timedependent increase serum concentration of proinflammatory cytokines in chloroform intoxicated rats compared to controls. Several studies that have published on this topic support this finding by reporting a similar elevation in pro-inflammatory cytokine response during exposure to chemical toxicants [38-41]. From the above, it is clear that damage caused by impaired liver function can lead to disruption of the body's health system as a result of immune suppression [42].



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Conclusions

This experimental investigation concluded the destructive effect of chloroform on liver function enzymes in the serum of a rat model. It also confirmed the immunotoxicity through the induction of pro-inflammatory cytokine response in a time-dependent increase.

Reference

- [1] Abakpa, A. M., Japhet, T., Omale, P., Chiyam, B. B., Peter, I., & Abu, G. O. (2024). Volatile Organic Compound in the Environment: Sources, Exposure and Mitigation. African Multidisciplinary Journal of Sciences and Artificial Intelligence, 1(1), 356-384.
- [2] Huang, L., Sang, C. N., & Desai, M. S. (2017). Beyond ether and chloroform—a major breakthrough with halothane. Journal of anesthesia history, 3(3), 87-102.
- [3] Cui, S. Q., Tao, Y. M., Jian, T. Z., Han, J., Ren, Y. L., Zhang, & Jian, X. D. (2022). An incident of chloroform poisoning on a university campus. World journal of emergency medicine, 13(2), 155.
- [4] Ali, Q. A., & Ghareeb, O. A. (2023). Drinking Water Quality and Its Impact on Public Health. Academia Repository, 4(9), 48-64.
- [5] Ali, Q. A., & Ghareeb, O. A. (2024). Proposed Solutions to Improve Deterioration of Drinking Water Quality. Global Scientific Review, 23, 34-46.
- [6] McCarty, P. L. (2010). Groundwater contamination by chlorinated solvents: history, remediation technologies and strategies. In situ remediation of chlorinated solvent plumes, 1-28.
- [7] Mezgebe, B., Palanisamy, K., Sorial, G. A., Sahle-Demessie, E., Aly Hassan, A., & Lu, J. (2018). Comparative study on the performance of anaerobic and aerobic biotrickling filter for removal of chloroform. Environmental engineering science, 35(5), 462-471.
- [8] Sekar, A., Varghese, G. K., & Varma, M. R. (2021). Chloroform—an emerging pollutant in the air. In New Trends in Emerging Environmental Contaminants (pp. 101-129). Singapore: Springer Singapore.
- [9] Fakhri, Y., Mohseni-Bandpei, A., Oliveri Conti, G., Keramati, H., Zandsalimi, Y., Amanidaz, N., & Baninameh, Z. (2017). Health risk assessment induced by chloroform content of the drinking water in Iran: systematic review. Toxin reviews, 36(4), 342-351.
- [10] Bruckner, J. V., Anand, S. S., & Warren, D. A. (2023). Toxic Effects of Solvents and Vapors: Introduction. Essentials of Toxicology, 2.
- [11] Ewaid, S. H., Abed, S. A., & Al-Ansari, N. (2020). Acute toxicity of the water chlorination byproduct (chloroform) in male mice. In AIP Conference Proceedings 2290, (1).
- [12] Hume, A. S., & Ho, K. (2019). Toxicity of solvents. In Basic Environmental Toxicology (pp. 157-184). CRC Press.
- [13] Lionte, C. (2010). Lethal complications after poisoning with chloroform—case report and literature review. Human & experimental toxicology, 29(7), 615-622.
- [14] Aguwa, U. S., Nnamdi, O. S., Nnabuihe, E. D., Elizabeth, E. C., Ogechi, A., Nzube, O. B., & Chijioke, O. (2020). Evaluating the effect of chloroform inhalation as a method of euthanasia on the cerebellum and hippocampus of adult wistar rats. Journal of Advances in Medical and Pharmaceutical Sciences, 22(6), 14-25.
- [15] Burcham, P. C., & Burcham, P. C. (2014). Target-organ toxicity: liver and kidney. An Introduction to Toxicology, 151-187.
- [16] Ghareeb, O. A., & Ali, Q. A. (2024). Pathological Disorders Caused by Atmospheric Nanoparticles. The Peerian Journal, 26, 44-51.
- [17] Ghareeb, O. A., & Ali, Q. A. (2024). Hepatotoxicity Induced by Some Metal Nanoparticles In Vivo. Global Scientific Review, 23, 25-33.
- [18] Bruckner, J. V., Anand, S. S., & Warren, D. A. (2023). Toxic Effects of Solvents and Vapors: Introduction. Essentials of



SEEJPH 2024 Posted: 16-08-2024

Toxicology, 2.

- [19] Li, Y., Cui, J., & Jia, J. (2021). The activation of procarcinogens by CYP1A1/1B1 and related chemo-preventive agents: a review. Current Cancer Drug Targets, 21(1), 21-54.
- [20] Wahlang, B., Hardesty, J. E., Head, K. Z., Jin, J., Falkner, K. C., & Beier, J. I. (2020). Hepatic injury caused by the environmental toxicant vinyl chloride is sex-dependent in mice. Toxicological Sciences, 174(1), 79-91.
- [21] Olowofolahan, A. O., & Olorunsogo, O. O. (2018). Induction of apoptosis in rat liver cells via caspase activation by chloroform fraction of methanol extract of drymaria cordata. Eur J Biomed Pharm Sci, 5(4), 73-83.
- [22] Priya, T., & Mishra, B. K. (2017). Enzyme mediated chloroform biotransformation and quantitative cancer risk analysis of trihalomethanes exposure in South East Asia. Exposure and Health, 9, 61-75.
- [23] Ghareeb, O. A., & Ali, Q. A. (2023). Pathotoxic Impact of Zinc Oxide Nanoparticles on Liver Function and Protective Role of Silymarin. Current Innovations in Disease and Health Research, 3, 153-161.
- [24] Rivera-Torruco, G., Muench, M. O., & Valle-Rios, R. (2024). Exploring extramedullary hematopoiesis: unraveling the hematopoietic microenvironments. Frontiers in Hematology, 3, 1371823.
- [25] Ghareeb, O. A., Ali, Q. A., Ramadhan, S. A., & Sultan, A. I. (2024). Concepts of One Health Approach and Achieving Sustainable Development. European Journal Pharmaceutical and Medical Research, 11(4), 383-389.
- [26] Naik, S. R., Gamare, D., & Bhopatrao, A. (2024). Chemical health hazards and toxicity of environmental pollutants on humans, animals and others: An overview. Journal of Toxicological Studies, 2(1), 1135-1135.
- [27] Parida, L., & Patel, T. N. (2023). Systemic impact of heavy metals and their role in cancer development: a review. Environmental Monitoring and Assessment, 195(6), 766.
- [28] Ghareeb, O. A. (2023). Adverse impact of titanium dioxide nanoparticles on hepato-renal functions and improved role of Rosmarinus officinalis. Journal of Natural Science, Biology and Medicine, 14 (1), 33-38.
- [29] Taher, G. N., Abdulla, G. M., & Ghareeb, O. A. (2024). Evaluation of some biochemical indicators in patients with gallstones. World Journal of Pharmaceutical and Medical Research, 10(7), 21-25.
- [30] Gryshchenko, V. A., Sysolyatin, S. V., & Gulevata, J. V. (2018). Phospholipid composition of blood plasma and internal organs of rats with diclofenac-induced hepatitis. Ukrainian Journal of Ecology, 8(3), 235-240.
- [31] Chima, U. O. P., Obianuju, O. R., & Ben, O. M. (2022). The Effect of Ethanol Leaf Extract of Rauwolfia vomitoria on Hepatic Markers of Chloroform Intoxicated Albino Wistar Rats. IAA Journal of Applied Sciences. 8 (1), 76-86.
- [32] Okechukwu, P. U., Nzubechukwu, E., Ogbansh, M. E., Ezeani, N., Nworie, M. O., & Ezugwu, A. (2015). The Effect of ethanol leaf extract of Jatropha curcas on chloroform induced hepatotoxicity in Albino rats. Global J Biotech & Biochem, 10, 11-15.
- [33] Somade, O. T., Ugbaja, R. N., Alli, A. A., Odubote, O. T., Yusuf, T. S., & Busari, B. T. (2018). Diallyl disulfide, an organo-sulfur compound in garlic and onion attenuates trichloromethane-induced hepatic oxidative stress, activation of NFkB and apoptosis in rats. Journal of Nutrition & Intermediary Metabolism, 13, 10-19.
- [34] Mehendale, H. M. (2017). Biochemical mechanisms of biphasic dose-response relationships: role of hormesis. In Biological Effects of Low Level Exposures to Chemical and Radiation, 59-94. CRC Press.
- [35] Das, S. (2020). Toxic gases. In Toxicology Cases for the Clinical and Forensic Laboratory (pp. 387-396). Academic Press.
- [36] Hobson, S. T., Richieri, R. A., & Parseghian, M. H. (2021). Phosgene: toxicology, animal models, and medical countermeasures. Toxicology mechanisms and methods, 31(4), 293-307.
- [37] Basij, M., Talebi, K., Ghadamyari, M., Hosseininaveh, V., & Salami, S. A. (2017). Status of resistance of Bemisia tabaci (Hemiptera: Aleyrodidae) to neonicotinoids in Iran and detoxification by cytochrome P450-dependent monooxygenases. Neotropical Entomology, 46, 115-124.
- [38] Somade, O. T., Ajayi, B. O., Olunaike, O. E., & Jimoh, L. A. (2020). Hepatic oxidative stress, up-regulation of proinflammatory cytokines, apoptotic and oncogenic markers following 2-methoxyethanol administrations in rats. Biochemistry and biophysics reports, 24, 100806.



SEEJPH 2024 Posted: 16-08-2024

- [39] Alam, R. T., Zeid, E. H. A., & Imam, T. S. (2017). Protective role of quercetin against hematotoxic and immunotoxic effects of furan in rats. Environmental Science and Pollution Research, 24, 3780-3789.
- [40] Lone, M. I., Nazam, N., Hussain, A., Singh, S. K., Dar, A. H., Najar, R. A., ... & Ahmad, W. (2016). Genotoxicity and immunotoxic effects of 1, 2-dichloroethane in Wistar rats. Journal of Environmental Science and Health, Part C, 34(3), 169-186.
- [41] Zabrodskii, P. F. (2018). The effect of chronic intoxication with 2-chloroethanol on immune responses, function of Th1 and Th2 lymphocytes and blood cytokine concentrations. Pharm Pharmacol Int J, 6(2), 00151.
- [42] Wang, H., Mehal, W., Nagy, L. E., & Rotman, Y. (2021). Immunological mechanisms and therapeutic targets of fatty liver diseases. Cellular & Molecular Immunology, 18(1), 73-91..