

HEPATOPROTECTIVE ACTIVITY OF POLYHERBAL EXTRACT AGAINST CCL4 INDUCED LIVER DAMAGE IN RATS

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KEYWORDS

ABSTRACT

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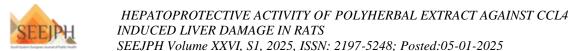
E. crassipes, CCl₄, hepatoprotective, hepatotoxicity, histopathology The current work evaluated the hepatoprotective effect of ethanolic extracts of *Eichhornia crassipes, Ipomoea aquatica*, and *Hydrilla verticillate* using silymarin against the CCl₄-induced in-vivo rat model at varying doses. Histological investigations were used to confirm the hepatoprotective action of EC, IP & HV, which was assessed by assessing serum biomarker levels. Rat's livers were effectively shielded from the damage caused by CCl₄ by the plant extract. Serum levels of bilirubin, total protein, alkaline phosphatase, serum glutamic-pyruvic transaminase and serum glutamic oxaloacetic transaminase were significantly decreased. In contrast to silymarin, the histological analysis showed a notable repair of liver structure. The traditional usage of polyherbs to comprehend its hepatoprotective potential has been scientifically validated by this investigation.

INTRODUCTION

Metabolic and histological abnormalities i.e., vascular lesions, granulomas, cholestasis, hepatitis, and zonal necrosis are characteristics of hepatotoxicity, or liver damage, which accounts for 5% of all injuries and is hence common. As a result, over a million deaths are recorded annually, with hepatocellular cancer or liver deformity being the indirect cause [1]. We are protected against a variety of internal and external illnesses by conventional or synthetic medications. Unfortunately, the medications used to treat liver problems are inadequate since they might have major long-term negative effects and do not completely protect the organ [2–5]. Understanding the function of complementary and alternative medicine as a treatment option for liver disease is thus essential [6,7]. Because of its abundance of polyphenols, CAM is known to have good therapeutic effectiveness with few negative side effects [8, 9]. Therefore, foods high in polyphenols may reduce the risk of chronic illnesses [5, 10-14].

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A monocotyledonous aquatic plant that floats freely, Eichhornia crassipes is a member of the Pontederiaceae family [15][16]. It has been used in wastewater treatment due to its ability to grow in polluted water and absorb heavy metals [17]. Both biofuels and biofertilizers are made from it. The plant has historically been used to treat intestinal worms, diarrhoea, vomiting, gas, and digestive problems [18]. Among the main metabolites that have been found from the different plant components are cellulose, hemicellulose, glycolipids, triacylglycerols, and heteropolysaccharides [19] [20]. Ponds, marshes, and other aquatic environments are home to its growth [21]. The phytochemical composition of several unusual green vegetables greatly inhibits glycoside hydrolases, including α -amylase and α -glucosidase [22][23].

Ipomoea aquatica or water spinach belongs to the Convolvulaceae family. It is also known as water spinach, river spinach, water morning glory, water convolvulus. It flourishes naturally in water which requires little care which commonly found in the banks of streams, ponds, ditches, lakes and marshes that usually floats or creeps along the shorelines but it can also climb over emergent vegetation, some also thrives in the water. It is used traditionally for the treatment of jaundice and liver diseases. The antioxidant property of water spinach is said to be due by the phytochemical, Flavonoid. [24].

Hydrilla verticillate is an aquatic plant with rapid growth and widespread dispersal. Particularly as water weeds, it may create problems in the water [25][26]. However, hydrilla plants have is a source of biomass for composting due to their vegetative reproductive systems (rhizomes and tubers), which may act as a place for reserves of carbohydrates [27]. Hydrilla verticillata is an aquatic plant that may act as a carbon source when it is fresh. For growth and development, plants need large quantities of the macroelements potassium, phosphorus, and nitrogen [28]. Denitrification is the process by which nitrogen gas is reduced during Hydrilla verticillata's breakdown. Waste will break down more quickly with bacterially aided phytoremediation [29]. In addition to the high expense of inorganic fertilizers and soil degradation caused by excessive use of chemical fertilizers, hydrilla plants may help address problems with natural resources that have not been fully used.

Hydrilla verticillata has a number of active compounds with antioxidant qualities. *Hydrilla verticillate* contains steroid [30]. The purpose of the research was to examine the hepatoprotective potential of ethanolic extract of polyherbs (EC IP & HV) on rats' liver damage caused by carbon tetrachloride.

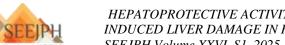
MATERIAL AND METHODS

Plant collection and authentication

The plant species Eichhornia crassipes, Ipomoea aquatica, and Hydrilla verticillate were procured from Patna, Bihar, and authenticated by professionals in the Department of ICAR- National Bureau of plant genetic resources, Pusa campus, Delhi. On the basis of visual observations and detail of the specimen it was identified by Scientist/Botanist R.K. Pamarthi and authenticated, and authentication certificated had been issued (ref. no. AC-137/2023, AC-136/2023, and AC-138/2023 respectively).

Extraction of the plant

These were washed thoroughly and dried in shade. The dried plant materials were crushed into coarse powder and then extracted through Soxhlet using ethanol. After extraction, the obtained slurry was filtered with filter paper and dried using rotatory evaporator [31].



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Animals and care

In this investigation, adult Wistar rats weighing 200-250g (either sex) were used. The Central animal facility at the All India Institute of Medical Sciences, New Delhi is where the animals were bought. With eight animals per cage, the animals were kept in normal polypropylene cages with bedding made of rice husk and standard climatic conditions (26±3°C, 65±1% RH, and 12-hour dark-light cycle. They were given regular pellet feed and distilled water as they acclimated for one to two weeks. The laws protecting animals used for scientific research were followed while conducting the animal studies.

Ethical statement

All experiments were conducted in rigorous compliance with established protocols. Ethical approval was secured from Innovative College of Pharmacy, Plot no. 6, Knowledge Park-2, Greater Noida U.P. India IAEC (1346/PO/Re/s/10CPCSEA) [32-34].

Acute toxicity study

The doses of extract utilized in this study were determined based on our prior findings from the acute toxicity assessment of the respective plants. The acute toxicity assessment was conducted in accordance with OECD guidelines 423. Administration of EV IP &HV at 2000mg/kg did not result in death of any animals. Therefore 250, 500 & 750mg/kg, 100, 200 & 400mg/kg & 50,75 & 100mg/kg respectively were chosen dose.

Experimental design and drug administration

Rodents were kept separately in 6 groups 4th, 5th & 6th group divided in 3 subgroups (12) (n= 8) as follows.

Group I (control) given normal diet.

Group II (disease control) given CCl₄ 1ml/kg. i.p. (mixed in olive oil 1:1) 2 times in 3 weeks) [11]. Group III (CCl₄ & Sily group) given CCl₄ 1ml/kg, i.p. (mixed olive oil 1:1) + silymarin (50mg/kg,

orally) for 21 days.

Group IV-VI given CCl₄ 1ml/kg, i.p. (olive oil 1:1) and at different doses of ethanolic extracts of all 3 plants orally (250, 500 & 750mg/kg, 100, 200 & 400mg/kg & 50,75 & 100mg/kg) for 3 weeks [35].

Preparation of liver tissue samples

All rats were fasted for 12 hours, were weighed and euthanized under proper anaesthesia using a combination of 0.15 ml/100 g xylazine (30 mg/kg BW) and 0.3 ml/100 g i.p ketamine (300 mg/kg BW) as proposed by Committee for the Purpose of Control and Supervision of Experiments on Animals (CCSEA). The blood was collected by retro-orbital puncture in an EDTA containing tube from all the experimental rats. The livers of each rat were removed, weighed and perfused in the ice-cold phosphate buffer of pH 7.0. A portion of the liver was preserved in 10% formaldehyde solution for histopathological evaluation.

Biochemical assays in blood

Alanine amino transaminase, aspartate amino transaminase, and alkaline phosphatase were quantified using an automated analyzer, alongside various liver markers such as total protein and albumin, adhering to established standard procedures [36].

Histopathological studies

A little section of the liver was removed, cleaned with regular saline, and prepared independently for histopathological analysis. The liver tissues were first preserved for at least 48 hours in 10% buffered



neutral formalin, then gradually dried in ethanol (50–100%), cleaned in xylene, and finally embedded in paraffin. A microtome was used to create the 4 μm slices. The liver slices were then rinsed with distilled water for five minutes after being dewaxed in xylene and rehydrated in a succession of alcohol grades. The liver slices were counterstained with acidic stain eosin for 20 s (H-E) dye after being stained for 40 s with basic stain hematoxylin [37]. Using an Olympus microscope, the sections were viewed at 400X magnification to check for any histopathological alterations, such as vacuolation, fatty changes, or necrosis of the cells.

Statistical analysis

The data are presented as mean \pm SE. A one-way ANOVA was conducted to compare different groups, followed by Tukey's multiple comparison tests using GraphPad Prism. A p-value of <0.05, p<0.01, and p<0.001 was taken as statistically significant.

RESULTS AND DISCUSSION

% Yield

Percentage yield of EV IP & HV extract isolated was found to be 18.35% 21.6 & 20.01% w/w.

Effect of ethanolic extract of EC HV & IP on biochemical parameters

Effect of ethanolic extract of EC HV & IP on the indices of hepatotoxicity based on liver marker enzymes

Administration of CCl4 markedly increased the levels of serum hepatic enzymes such as ALT, AST and ALP in the disease-induced group as compared to the control group (Table 1). The increase in these enzyme activities was significantly decreased with the treatment with extract of EV IP & HV. The high concentration group of EV IP & HV showing better results than the low concentration. These findings suggest a dose-dependent response for the EC and IP extracts, while the HV extracts exhibit less consistent effectiveness in reducing ALT, AST and ALP levels.

Table 1. Effect of plant extract on ALP ALT &AST level in CCl4 induced hepatoxic animals

Treatment	ALP (U/L) level	AST (U/L) level	ALT (U/L) level
Normal saline	49.31± 3.82	13.65± 1.22	18.1±1.276
CCl ₄ (1ml/kg, i. p.)	130.2± 6.76	91.08± 2.90	54.05±2.901
Silymarin (50mg/kg) + CCl4	52.77± 3.64	17.38± 1.21	21.73±1.214
EC 250 mg/kg + CCl4	81.67± 6.26	44.81± 2.08	36.77±2.08
EC 500 mg/kg + CCl4	71.86± 4.51	39.75± 1.50	28.09±1.50
EC 750 mg/kg + CCl4	60.06± 5.54	36.13± 1.50	24.16±1.84
IP 100 mg/kg + CCl4	101.4± 6.43	50.53± 2.14	42.49±2.14
IP 200 mg/kg + CCl4	98.84± 7.92	46.91± 2.64	38.87±2.64
IP 400 mg/kg + CCl4	80.03± 5.18	41.35± 1.91	30.81±1.72
HV 50 mg/kg + CCl4	105.4± 6.43	69.51± 2.44	43.85±2.14
HV 75 mg/kg + CCl4	97.45± 5.63	64.35± 2.40	41.19±1.87
HV 100 mg/kg + CCl4	101.6± 6.27	57.4± 2.09	37.99±2.09



EC - Eichhornia crassipes, IP - Ipomoea aquatica, HV - Hydrilla Verticillate

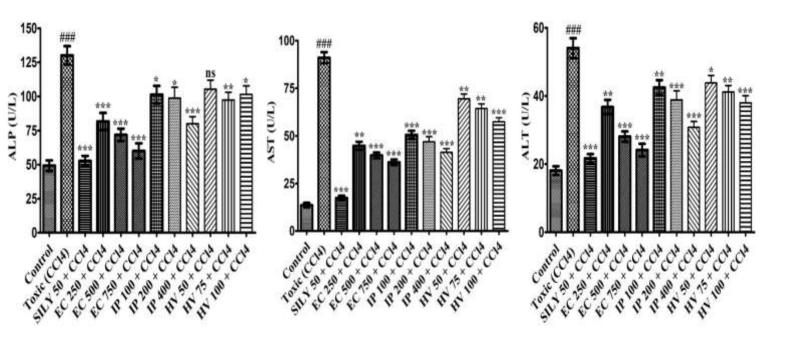


Figure 1 Level of ALP AST ALT level in animals
Data are presented as mean \pm SD, with significant differences indicated at *p<0.05, **p<0.01, and
***p<0.001 when compared to the control group
Effect of ethanolic extract of EC HV & IP on total protein and albumin level

As shown in Table 2, CCl4 administration decreased the albumin (to 1.618 g/dL,) and total protein level (to 3.268 g/dL) as compared to the control. The albumin level was increased on treatment with extract of EV IP & HV at a high dose and silymarin (4.001 g/dL) significantly increased the albumin to 3.356 g/dL, 3.622 g/dL, and 3.803 g/dL, respectively, as compared to the control (4.683 g/dL). The total protein level was increased on treatment with extract of EV IP & HV at a high dose and silymarin (6.601 g/dL) significantly increased the albumin to 6.265 g/dL, 5.01 g/dL, and 4.825 g/dL, respectively, as compared to the control (6.783 g/dL). Treatments with Silymarin, as well as various doses of EC and IP extracts, demonstrate potential in increasing albumin & total protein levels and restoring liver function. The findings suggest a dose-dependent response for the EC and IP extracts, while the HV extracts exhibit less consistent effectiveness in increasing albumin & total protein levels.

Table 2. Effect of plant extract on albumin & Total Protein level in CCl4 induced hepatoxic animals

Treatment	Total protein (mg//dL) level	Albumin (g/dl)
Normal saline	6.78±0.42	4.68± 0.25
CCl ₄ (1ml/kg, i. p.)	3.26±0.31	1.61± 0.15
Silymarin (50mg/kg) + CCl4	6.60±0.28	4.00± 0.30
EC 250 mg/kg + CCl4	5.76±0.35	3.35 ± 0.26



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EC 500 mg/kg + CCl4	5.97±0.36	3.62 ± 0.30
EC 750 mg/kg + CCl4	6.26±0.26	3.80 ± 0.26
IP 100 mg/kg + CCl4	5.45±0.20	3.00± 0.09
IP 200 mg/kg + CCl4	5.30±0.34	3.20 ± 0.34
IP 400 mg/kg + CCl4	5.01±0.38	3.74 ± 0.30
HV 50 mg/kg + CCl4	3.93±0.23	2.16± 0.17
HV 75 mg/kg + CCl4	4.42±0.29	2.50± 0.19
HV 100 mg/kg + CCl4	4.82±0.29	2.85 ± 0.22

EC - Eichhornia crassipes, IP - Ipomoea aquatica, HV - Hydrilla Verticillate

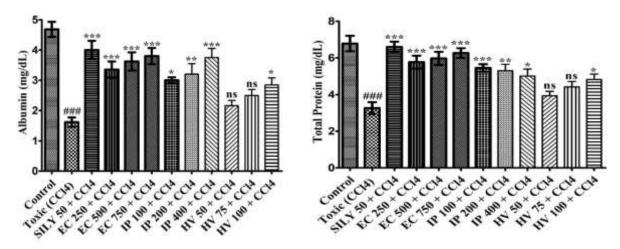


Figure 2 Albumin & Total protein level in animals Data are presented as mean \pm SD, with significant differences indicated at *p<0.05, **p<0.01, and ***p<0.001 when compared to the control group

Effect of ethanolic extract of EC HV & IP on bilirubin level

The administration of CCl4 elevated the levels of serum bilirubin (0.1848 mg/dL) as compared to the control group (Table 3). After the treatment with ethanolic extract of EC HV & IP at higher dose bilirubin was lessened to (0.2837, 0.7589 & 1.243 mg/dL) respectively and 0.2298 mg/dL (with silymarin) as compared to control (0.1848 mg/dL). These findings suggest a dose-dependent response for the EC and IP extracts, while the HV extracts exhibit less consistent effectiveness in reducing bilirubin levels.

Table 3. Effect of plant extract on bilirubin level in CCl4 induced hepatoxic animals

Treatment	Bilirubin (mg//dL) level	
Normal saline	0.18±0.04	
CCl ₄ (1ml/kg, i. p.)	1.9±0.07	
Silymarin (50mg/kg) + CCl4	0.22±0.02	
EC 250 mg/kg + CCl4	0.35±0.04	
EC 500 mg/kg + CCl4	0.32±0.03	
EC 750 mg/kg + CCl4	0.28±0.07	
IP 100 mg/kg + CCl4	0.96±0.07	



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IP 200 mg/kg + CCl4	0.87±0.04	
IP 400 mg/kg + CCl4	0.75±0.10	
HV 50 mg/kg + CCl4	1.5±0.11	
HV 75 mg/kg + CCl4	1.3±0.15	
HV 100 mg/kg + CCl4	1.2±0.13	

EC - Eichhornia crassipes, IP - Ipomoea aquatica, HV - Hydrilla Verticillate

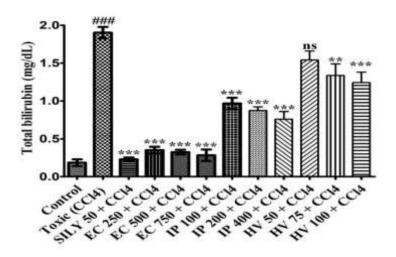


Figure 3. Bilirubin level in animals Data are presented as mean \pm SD, with significant differences indicated at *p<0.05, **p<0.01, and ***p<0.001 when compared to the control group

Table 4. Effect of plant extract on liver weight to body weight ratio of animals

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Groups	Body weight	Liver weight	Ratio		
Control	227.48±11.61	4.69±0.88	0.020		
CCl ₄ (1ml/kg, i. p.)	177.82±7.64	7.76±0.88	0.004		
Silymarin (50mg/kg) + CCl4	219.17±7.37	4.59±0.88	0.021		
EC 250 mg/kg+ CCl4	185.66±9.39	5.72±0.88	0.031		
EC 500 mg/kg+ CCl4	189.02±7.64	5.58±0.89	0.029		
EC 750 mg/kg+ CCl4	194.87±5.87	5.48±0.88	0.028		
IP 100 mg/kg+ CCl4	191.59±7.39	5.45±0.89	0.028		
IP 200 mg/kg+ CC14	196.00±6.36	5.16±0.88	0.026		
IP 400 mg/kg+ CCl4	200.13±0.28	4.96±0.88	0.024		
HV 50 mg/kg+ CCl4	199.08±5.23	4.02±0.88	0.020		
HV 75 mg/kg+ CCl4	205.42±7.45	4.14±0.54	0.020		
HV 100 mg/kg+ CCl4	210.21±4.48	4.01±0.88	0.019		

EC - Eichhornia crassipes, IP - Ipomoea aquatica, HV - Hydrilla Verticillate

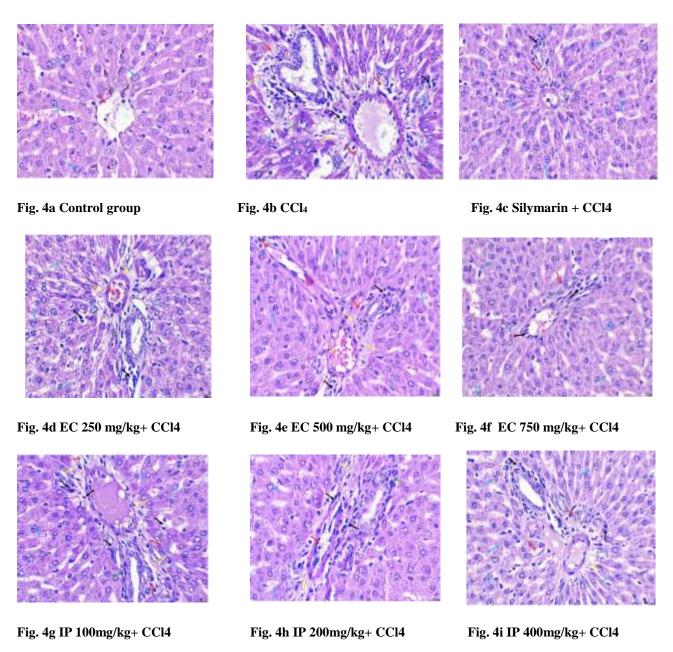
As table 4 provides data on the liver weight to body weight ratio for animals in different treatment groups, which is a key metric for assessing the effects of plant extracts on liver health. This ratio helps evaluate liver hypertrophy or damage by comparing liver size relative to body weight. A higher ratio could indicate liver enlargement, which may be a response to toxicity, while a lower ratio could suggest liver shrinkage or other forms of liver dysfunction.

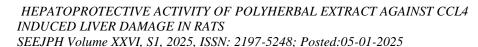


Histopathological evaluation of the rat livers (Table 4; Fig. 4) showed that the hepatocytes of a healthy rat (from the control group) had a normal architecture (Fig. 4a), whereas, in contrast, CCl4 induced severe hepatocyte necrosis, inflammation, and hemorrhage (Fig. 4b). After treatment with EV IP & HV, the severity of CCl4-induced liver intoxication was reduced in a dose-dependent manner (Fig. 4d,e,f and 4g,h,i and 4j,k,l), although the treatment with silymarin showed better result (Fig. 4c).

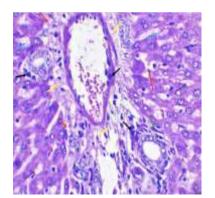
The table highlights how different plant extracts at varying dosages can influence the liver weight to body weight ratio, indicating that some extracts (like EC) may increase liver size, while others (like HV) might reduce it or have little effect compared to the control. These differences are important for understanding the potential liver-protective or liver-modifying properties of the extracts tested.

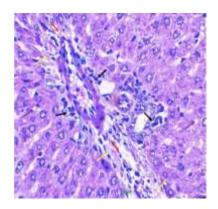
Figure 4 Histopathology of plant extract treated CCL4-induced hepatotoxic rats











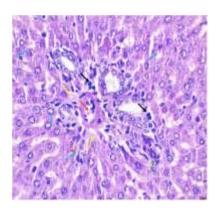


Fig.j HV 50 mg/kg+ CCl4

Fig.k HV 75 mg/kg+ CCl4

Fig. 41 HV 100 mg/kg+ CCl4

Toxic group shows significantly damaged hepatocytes (blue arrow). High cellular density, dense cytoplasm, and significant fatty changes (yellow arrow) and vacuolation (purple arrow) are seen. Marked cellular disintegration (red arrow), pyknosis (black arrow), inflammatory infiltrate (green arrow) are seen. The histomorphological characteristics manifest marked hepatic toxicity. (H&E, 400X). Std. group shows minimal histological damage to liver tissue. No congestion in the central vein (CV) is seen. The section shows normal and trabecular-arranged hepatocytes, which are polygonal in shape (blue arrow). No fatty changes, non-dilated sinusoids, well-placed Kuffer's cells, no lipoid vacuoles, and minimal cellular disintegration (red arrow) along with mild pyknosis (black arrow) and no evidence of inflammatory infiltration were seen (H&E, 400X). EC 250 shows moderate histological damage to liver tissue. Moderate congestion in the central vein (CV) is seen (red arrow). The section shows mildly damaged hepatocytes (blue arrow). Moderate fatty changes (yellow arrow), non-dilated sinusoids, well-placed Kuffer's cells, mild lipoid vacuoles (white arrow) and severe cellular disintegration along with mild pyknosis (black arrow) and mild inflammatory cells (green arrow) were seen (H&E, 400X). EC 500 shows minimal histological damage to liver tissue. Mild congestion in the central vein (CV) is seen (orange arrow). The section shows normal and trabecular-arranged hepatocytes, which are polygonal in shape (blue arrow). Minimal fatty changes (yellow arrow), non-dilated sinusoids, well-placed Kuffer's cells, minimal lipoid vacuoles (white arrow), and minimal cellular disintegration (red arrow) along with mild pyknosis (black arrow) and no evidence of inflammatory infiltration were seen (H&E, 400X). EC 750 shows minimal histological damage to liver tissue. No congestion in the central vein (CV) is seen. The section shows normal and trabecular-arranged hepatocytes, which are polygonal in shape (blue arrow). No fatty changes, non-dilated sinusoids, well-placed Kuffer's cells, no lipoid vacuoles, and minimal cellular disintegration (red arrow) along with mild pyknosis (black arrow) and no evidence of inflammatory infiltration were seen (H&E, 400X). IP 100 shows significantly damaged hepatocytes (blue arrow). High cellular density, dense cytoplasm, and significant fatty changes (yellow arrow) and vacuolation (purple arrow) are seen. Marked cellular disintegration (red arrow), pyknosis (black arrow), inflammatory infiltrate (green arrow) are seen. The histomorphological characteristics manifest marked hepatic toxicity. (H&E, 400X). IP 200 shows moderate histological damage to liver tissue. Moderate congestion in the central vein (CV) is seen (red arrow). The section shows mildly damaged hepatocytes (blue arrow). Moderate fatty changes (yellow arrow), non-dilated sinusoids, well-placed Kuffer's cells, mild lipoid vacuoles (white arrow) and severe cellular disintegration along with mild pyknosis (black arrow) and mild inflammatory cells (green arrow) were seen (H&E, 400X). IP 400 shows mild histological damage to liver tissue. No congestion in the central vein (CV) is seen. The section shows normal and trabecular-arranged hepatocytes, which are polygonal in shape (blue arrow). Minimal fatty changes (yellow arrow), non-dilated sinusoids, well-placed Kuffer's cells, no lipoid vacuoles (white arrow), and minimal cellular disintegration (red arrow) along with mild pyknosis (black arrow) and no evidence of inflammatory infiltration were seen (H&E, 400X). HV 50 shows significantly damaged hepatocytes (blue arrow). High cellular density, dense cytoplasm, and significant fatty changes (yellow arrow) and vacuolation (purple arrow) are seen. Marked cellular disintegration (red arrow), pyknosis (black arrow), and inflammatory infiltrate (green arrow) are seen. The histomorphological characteristics manifest marked hepatic toxicity. (H&E, 400X). HV 75 shows significantly damaged hepatocytes (blue arrow). High cellular density, dense cytoplasm, and



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significant fatty changes (yellow arrow) and vacuolation (purple arrow) are seen. Marked cellular disintegration (red arrow), pyknosis (black arrow), inflammatory infiltrate (green arrow) are seen. The histomorphological characteristics manifest marked hepatic toxicity. (H&E, 400X). HV 100 shows moderate histological damage to liver tissue. Moderate congestion in the central vein (CV) is seen (red arrow). The section shows mildly damaged hepatocytes (blue arrow). Moderate fatty changes (yellow arrow), non-dilated sinusoids, well-placed Kuffer's cells, mild lipoid vacuoles and severe cellular disintegration along with mild pyknosis (black arrow) and mild inflammatory cells (green arrow) were seen (H&E, 400X).

Conclusion

Liver damage can result in inflammation, fibrosis, and necrosis, ultimately leading to liver failure [38]. Various plants have been utilized for their medicinal properties across numerous regions globally [39– 42]. Silymarin, a highly potent phytochemical, is utilized in the treatment of hepatic diseases [34]. EP, classified within the polyphenol category, serves as a powerful antioxidant that combats ROS-induced oxidative stress, making it a valuable agent in the management of liver diseases [43]. CCl4 is frequently utilized in studies of liver toxicity involving experimental animal models, as it induces lipid peroxidation through the generation of free radicals [44]. CCl4 serves as an exemplary animal model known for its association with free radical-induced hepatotoxicity caused by xenobiotics [45]. This analysis demonstrates that CCl4 induced considerable hepatic damage and oxidative stress in animals, as indicated by changes in liver function tests and antioxidant enzyme levels [46, 47]. This study's findings demonstrated that CCl4 exposure led to hepatotoxicity and oxidative stress in rats. The intraperitoneal injection of CCl4 results in an increase in the activity of ALT, AST, and ALP, while simultaneously decreasing the levels of TP and serum albumin, as well as reducing the level of CYP450. The administration of EV IP & HV and SILY effectively mitigates all harmful effects observed in the livers of rats. These factors diminish oxidative stress, inhibit the infiltration of inflammatory cells, enhance the regenerative potential of injured tissues, and decrease liver apoptosis. Therefore, it is our assertion that incorporating natural products like EV IP & HV and SILY may help mitigate the toxic effects associated with exposure to xenobiotics, including CCl4 and other harmful substances.

In conclusion, the ethanolic extract has shown the capacity to preserve the liver's functioning state. The ethanolic extract of E. crassipes has been shown to be one of the herbal treatments for liver disease, according to the above early investigation.

Abbreviations

CCl4: Carbon tetrachloride

SILY: Silymarin

ALT: Alanine amino transaminase AST: Aspartate amino transaminase

ALP: Alkaline phosphatase

TP: Total protein

EV: Eichhornia crassipes IP: Ipomoea aquatica HV: Hydrilla verticillate CYP450: Cytochrome P450



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